

Evaluation of molnupiravir analogues as novel coronavirus (SARS-CoV-2) RNA-dependent RNA polymerase (RdRp) inhibitors - an *in silico* docking and ADMET simulation study

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ABSTRACT: The severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2), which is characterized by a wide range of symptoms including fever, dry cough, headache, decreased sense of taste and smell, was first identified in Wuhan, China in December 2019. Currently, the nucleoside analog, remdesivir has been approved for emergency use authorization (EUA) by the regulatory agencies for the treatment of COVID-19 patients. The need for new antiviral agents has been continuing due to the some disadvantages of remdesivir. Molnupiravir (MLN) that is developed for the treatment of hepatitis C virus (HCV), have been reported to show antiviral activity against SARS-CoV-2 according to the results of a high throughput screen of nucleoside analogs and also phase II/III clinical trials of MLN is ongoing. In this study, fifty four MLN analogs (twelve of them are found to be reported in the literature whereas forty two of them are novel molecules) against SARS-CoV-2 RdRp were designed and evaluated for their potential antiviral activity by using molecular modelling studies. While among the designed MLN analogs, compound **C17** was found to have the best potential inhibitor with -7.3 kcal/mol binding energy that is higher than molnupiravir and its active form EIDD-1931. Therefore, the isobutyric acid ester and monophosphate forms of **C17** were also compared to the related MLN derivatives in terms of active site interactions. Lastly, the ten compounds with the best binding affinity including **C17** were tested *in silico* for bioavailability, drug-likeness, ADME and safety profiles and were found to exhibit similar bioavailability and safety profile to MLN.

KEYWORDS: Molnupiravir; SARS-CoV-2 RdRp; ADMET prediction; docking studies.

1. INTRODUCTION

COVID-19 (human coronavirus) that is caused by coronavirus-2 (SARS-CoV-2), the cause of rapidly increasing numbers of severe pneumonia-like symptoms with fatal consequences, has spread rapidly around the world since December 2019. COVID-19 is the most disastrous disease in the recent years with more than one million deaths and millions infected, was declared a pandemic on 11th March, 2020 by WHO and since then, 233,503,524 confirmed cases of COVID-19, including 4,777,503 deaths were reported [1]. All viruses, including SARS-CoV-2, change over time and any new mutations can potentially increase or decrease infectiousness and virulence. However several vaccines are available against SARS-CoV-2 strain, the numerous new SARS-CoV-2 variants have been detected and some of these variants including SARS-CoV-2 Alpha, Beta, Gamma, Delta, and Lambda may increase the risk of possible re-infection or reduced effectiveness of vaccination [2]. Besides all this, considering the negative economic and social effects of this pandemic which has arisen due to the mandatory isolation and quarantine of millions of people, the need for the development of specific antiviral agents is increasing day by day [3]. As computer-aided drug discovery provides significant advantage for the prediction of potential molecules even before synthesis, preclinical testing and clinical trials in several years, *in silico* drug design and development studies increases which focuses the treatment of COVID-19, the disaster needed urgent solution.

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To date, seven different human coronavirus strains which of two are *alphacoronaviruses* (229E and NL63 strains) and others are *betacoronaviruses* (OC43, HKU1, SARS, MERS, and SARS-CoV-2 strains) have been reported [4]. SARS-CoV-2 is a member of *betacoronaviruses*, such as the most aggressive strains SARS and MERS human coronaviruses with their remarkable mortality rates (10% and 36%, respectively). SARS-CoV-2 genome encodes 16 non-structural proteins which are essential for the replication such as papain-like protease (nsp3), chymotrypsin-like main protease (3CL protease, nsp5), primase complex (nsp7-nsp8), RNA-dependent RNA polymerase RdRp (nsp12), helicase (nsp13), and exoribonuclease (nsp14) [3]. Among these viral enzymes, RdRp is a crucial one, presenting an attractive target for the development of new antiviral agents against SARS-CoV-2 due to its essential role in viral RNA synthesis in the life cycle of RNA viruses. Furthermore, inhibition of the enzyme is not expected to cause target-related side effects due to the absence of a functional equivalent to RdRp in mammalian cells. The development of effective RdRp inhibitors to block viral replication has long been a research topic for many scientists. There are two known classes of RdRp inhibitors: nucleoside analogue inhibitors (NIs) and non nucleoside analogue inhibitors (NNIs) [5]. A nucleoside analogue known as remdesivir remains as the only drug approved by regulatory agencies to treat COVID-19. Due to the difficulty of synthesis, intravenous administration and cost, and failure in some clinical trials for their beneficial effects, efforts have focused on the development of new alternative inhibitors of SARS-CoV-2 replication. It has been reported that the mutagenic effects of known antiviral drugs and their analogues on SARS-CoV-2 were examined in many studies in the literature (Elfiky, 2020; Esam, 2022). Among them, molnupiravir (MLN, Synonyms: EIDD-2801; MK-4482), a prodrug, is a promising new drug targeting the coronavirus RNA polymerase based on results of reported many studies [6–9]. MLN is currently in phase II/III clinical trials based on encouraging preclinical data and its lack of toxicity and severe adverse side effects in phase I clinical trials [10].

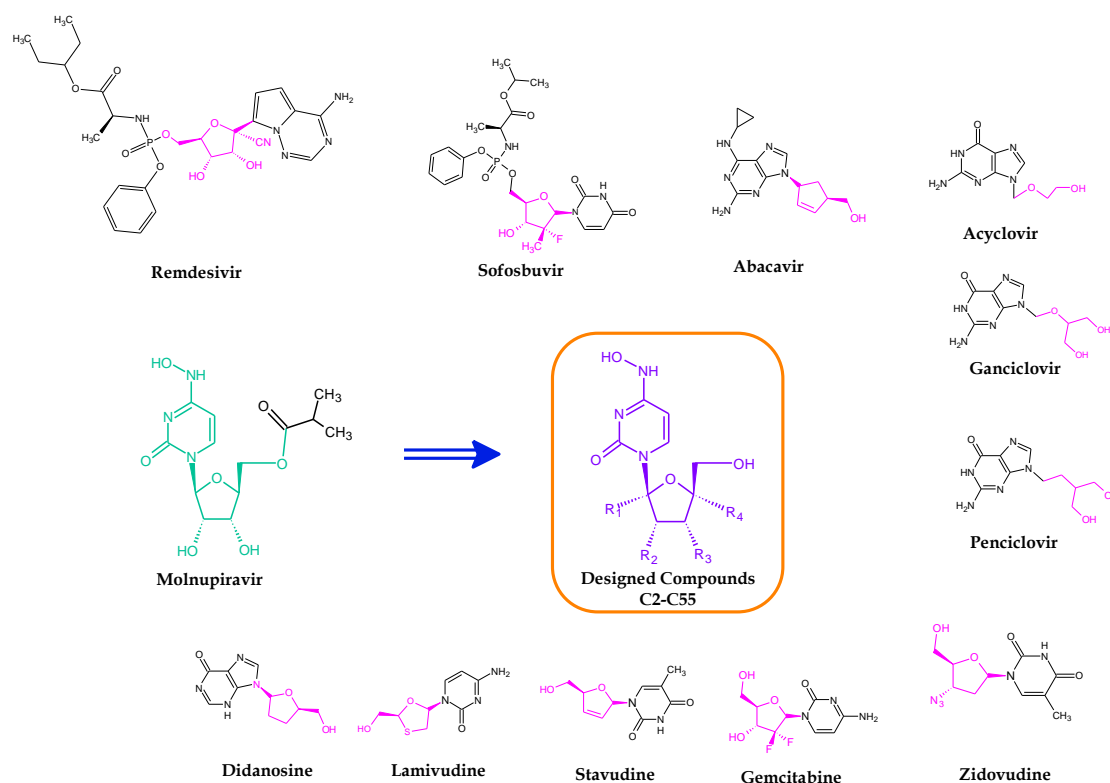


Figure 1. Design of the novel molnupiravir analogs from the known antiviral drugs.

Molnupiravir is a prodrug of β -(D)-N⁴-hydroxycytidine (NHC) which is known as EIDD-1931 (also called as compound **C1** in this article). The triphosphorylated derivative of EIDD-1931 is a substrate for viral RNA polymerases and interferes with viral replication. As known, cytidine is formed when cytosine is attached to a ribose ring and is a nucleoside molecule. The aim of this study was to design new analogs of molnupiravir analogs by making various deliberate modifications to the ribose ring of EIDD-1931 (Figure 1) and to evaluate the *in silico* inhibition potential of SARS-CoV-2 RdRp. The detailed ADME and safety profiles for the most active compounds identified in the screening were also calculated by *in silico* studies.

For this purpose, we designed fifty four molnupiravir analogue compounds **C2-C55** -twelve of them were reported as antiviral agents even compound **C13** was patented to using in anti-COVID-19 therapy by Lili and Shaochun [11-17]- with various modifications on the EIDD-1931 by using the structures of known antiviral drugs. In our study, the *in silico* screening results of the designed compounds and their estimated drug potentials are discussed in detail.

2. RESULTS and DISCUSSION

2.1. Molecular docking studies

Nucleoside analogs which inhibit RdRp, i.e. molnupiravir and gemcitabine act as pyrimidine pseudo-bases after phosphorylation and cause irreversible mutations in the viral genome or inhibit the viral RdRp directly, in a concentration-dependent manner. The presence of a ribose ring in the chemical structure of nucleoside analogs are used as alternative substrates for endogenous nucleotides, causing mutations on the RNA chain, but it seems not to be crucial for antiviral activity [18]. Therefore, while designing our compounds, besides the modifications on the ribose ring, we also examined the active site interactions of the straight and branched chains instead of the ribose ring, as in the cases of acyclovir, ganciclovir and penciclovir. Moreover, we also evaluated the active site interactions of the 1,3-oxathiolane and cyclopentene rings instead of the ribose ring, as lamivudine and abacavir, respectively. In our study, we used SARS-CoV-2 RdRp enzyme structure (6m71) the RNA strand free to evaluate the interactions of the designed MLN analogues (**C2-C55**) which has modifications on the ribose ring with the active site since known that nucleoside analogs RdRp inhibitors display mutagenic effect on the RNA strand after the phosphorylation.

The inhibition potentials of the designed and previously reported compounds (**C2-C55**) and molnupiravir against the SARS-CoV-2 RdRp enzyme were investigated using computer-aided methods. Compound **C13** has been reported as anti-COVID-19 agent better than MLN by *in vivo* studies [16]. Dissimilar the previous study the active site interactions of **C13** against SARS-CoV-2 RdRp were evaluated and discussed with the other designed compounds. When the results were evaluated, it was determined that all of the designed compounds showed quite significant affinity for binding to the active site of the enzyme and their binding energies were in the range of -7.3 to -4.8 kcal/mol (see Supporting Information, Table S1). As a result of *in silico* screening, the binding energies of the 10 compounds with the highest potential enzyme inhibition and their interactions with the active site are given in Table 1. Moreover, these selected compounds **C13-C15**, **C17-C19**, **C30**, **C32**, **C34** and **C48** have higher binding energy than MLN.

Firstly we examined the interactions of the active form of molnupiravir, **C1** with SARS-CoV-2 RdRp active site to compare with our designed compounds **C2-C55**. While β -D-N⁴-hydroxy group of cytidine showed two hydrogen bond with Thr556 residue as a donor, oxo group which is located at second position of cytidine ring, and nitrogen atom which is located at third position exhibited hydrogen bond interaction with Arg553 residue as an acceptor. Additionally, the hydroxyl group which is located at the fifth position of the ribose ring has two hydrogen bond interactions with Thr687 and Asn691 residues as donor and acceptor, respectively. Finally the distance between pyrimidine ring and Asp623 residue was found appropriate to pi-anion interaction (Figure 2D and 2E).

As a result of docking simulations, the best potential enzyme inhibitors were estimated as compounds **C13-C15** and **C17-C19** which are nucleoside analogues in which cyano group is introduced at 1' carbon of cytidine. Compound **C17** has been found to have the highest affinity with a greater number of active site interactions and a binding energy of -7.3 kcal/mol. These results are better than the values calculated for previously reported anti-COVID-19 agents, **C1** and **C13** (Table 1). Predicted conformation of compound **C17** superposed to **C1** in complex with SARS-CoV-2 RdRp is given in Figure 2A. When we examined those interactions between **C17** and the active site of SARS-CoV-2 RdRp, we detected both two hydrogen bond interactions between 3'-hydroxy group of cytidine and Tyr619, Lys621 residues, and halogen bond interaction between 2'-fluoro atom of cytidine and Lys621 residue different from compound **C1**. Furthermore, the pyrimidine ring has exhibited pi-anion and pi-cation interaction with Arg553 and Asp623 residues, respectively (Figure 2B and 2C). Similarly for the other derivatives which possess cyano group, while substitution of the 3th and/or 4th positions of the ribose ring with fluorine and/or hydroxyl groups resulted in increased activity, the binding energy to the active site was found to be decreased for compound **C16** which is a 3,4-dideoxy derivative (Table S1).

Table 1. Interactions of the selected compounds with the active site of SARS-CoV-2 RdRp enzyme.

Compounds	Binding Energy (kcal/mol)	H-bond interactions (Residue)	Distance (Å)
C1 (EIDD-2801)	-6.0	Arg553	2.24, 2.40 and 2.86
		Thr556	2.75 and 3.13
		Thr687	2.69
		Asn691	2.29
C13	-6.6	Arg555	3.13
		Asp623	4.37
C14	-6.4	Lys621	2.99
		Cys622	3.89
		Asp623	3.81
C15	-6.4	Lys621	3.08
		Cys622	3.86
		Asp623	3.85
		Thr680	4.44
C17	-7.3	Asp452	2.30
		Arg553	2.41 and 3.08
		Ala554	2.22
		Thr556	2.35
		Tyr619	2.54
		Lys621	2.75 and 2.55
C18	-6.5	Asp760	2.04
		Asp452	4.20
		Lys621	3.04
		Cys622	3.96
C19	-6.7	Asp623	3.84
		Lys621	3.04
		Asp623	3.84
C30	-6.4	Thr556	4.15
		Thr680	4.44
		Thr687	3.48
C32	-6.5	Thr556	3.46 and 3.51
		Thr556	3.91 and 3.94
C34	-6.3	Cys622	4.4
		Thr680	4.33
		Thr687	3.55
C48	-6.5	Lys621	3.04
		Asp623	3.92
C17_PD	-6.8	Asp452	2.27
		Ala554	2.27
		Thr556	2.39
MLN	-5.7	Arg553	2.64
		Thr556	2.62
		Asn691	2.50
		Ser759	2.25
		Asp760	2.45
C17_MP	-6.9	Arg553	1.58
		Ala554	2.31
		Tyr619	2.80
		Asp623	2.52
MLN_MP	-6.3	Arg553	2.32 and 2.85
		Thr556	2.56 and 3.42
		Asn691	2.66 and 2.96
		Ser759	2.58

Abbreviations: **MLN**: Molnupiravir, **C17-PD**: prodrug form of compound **C17**, **C17-MP**: monophosphate derivatives of compound **C17**, **MLN_MP**: monophosphate derivatives of Molnupiravir.

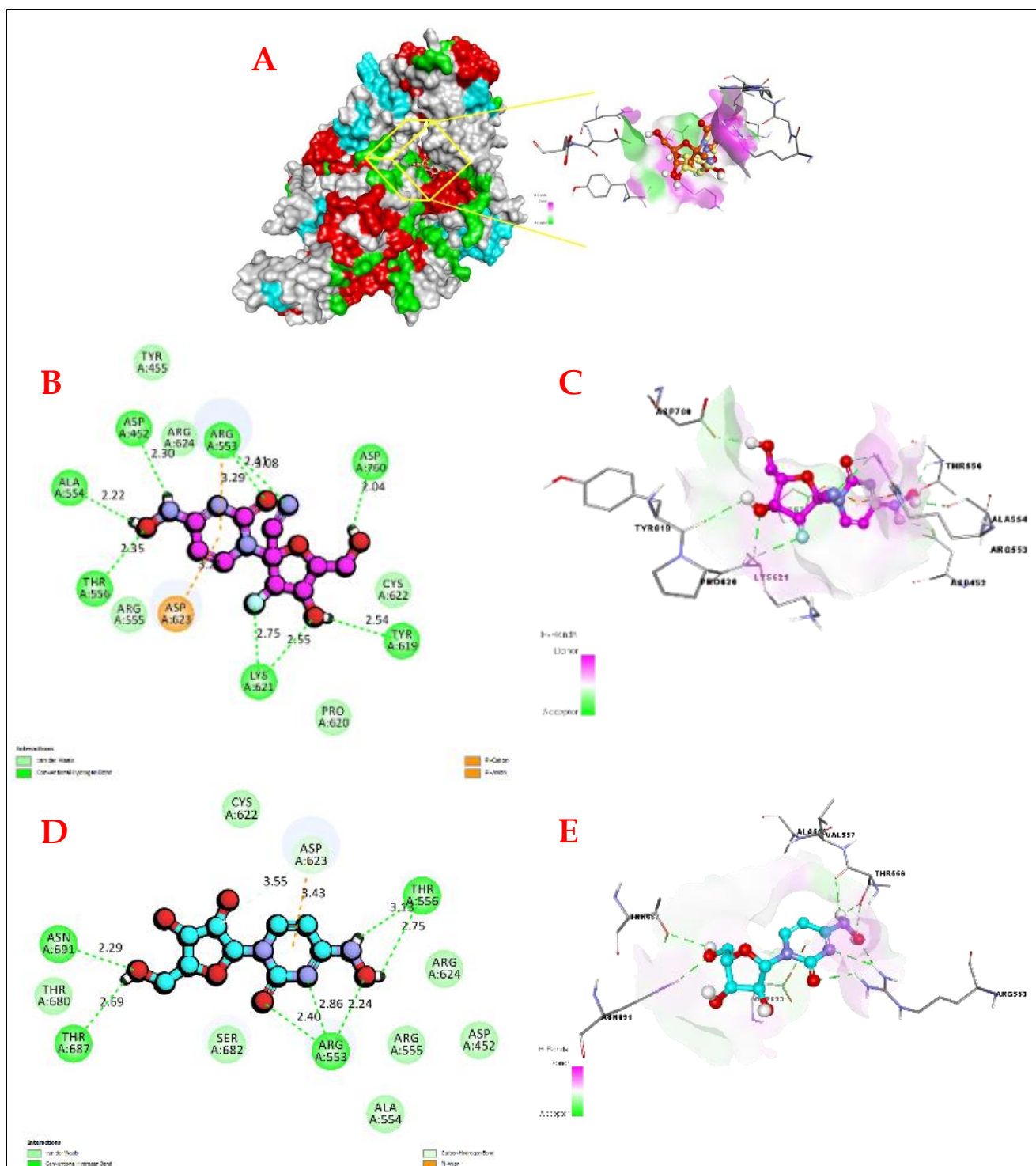


Figure 2. A) Predicted conformations of compound **C17** (orange) superposed to **C1** (yellow) in complex with SARS-CoV-2 RdRp. B) Interactions between compound **C17** and SARS-CoV-2 RdRp as 2D diagram. C) Binding orientation of compound **C17** in the active site of SARS-CoV-2 RdRp. D) Interactions between compound **C1** and SARS-CoV-2 RdRp as 2D diagram. E) Binding orientation of compound **C1** in the active site of SARS-CoV-2 RdRp.

As a result of docking simulations, the best potential enzyme inhibitors were estimated as compounds **C13-C15** and **C17-C19** which are nucleoside analogues in which cyano group is introduced at 1' carbon of cytidine. Compound **C17** has been found to have the highest affinity with a greater number of active site interactions and a binding energy of -7.3 kcal/mol. These results are better than the values calculated for previously reported anti-COVID-19 agents, **C1** and **C13** (Table 1). Predicted conformation of compound **C17**

superposed to **C1** in complex with SARS-CoV-2 RdRp is given in Figure 2A. When we examined those interactions between **C17** and the active site of SARS-CoV-2 RdRp, we detected both two hydrogen bond interactions between 3'-hydroxy group of cytidine and Tyr619, Lys621 residues, and halogen bond interaction between 2'-fluoro atom of cytidine and Lys621 residue different from compound **C1**. Furthermore, the pyrimidine ring has exhibited pi-anion and pi-cation interaction with Arg553 and Asp623 residues, respectively (Figure 2B and 2C). Similarly for the other derivatives which possess cyano group, while substitution of the 3th and/or 4th positions of the ribose ring with fluorine and/or hydroxyl groups resulted in increased activity, the binding energy to the active site was found to be decreased for compound **C16** which is a 3,4-dideoxy derivative (Table S1).

Among the most active compounds, **C30**, **C32** and **C34** contain an azido group on the 4' carbon of cytidine. While these compounds exhibited similar hydrogen bonding interactions to other designed cytidine derivatives, the azido group has been located in the distance that can make the charge transfer interaction with Asp760 residue which identifies the active site. This contribution increases the affinity into the active site. The *in silico* binding affinities of compounds **C39**, **C45** and **C48** decrease when the ribose ring is replaced by the 1,3-oxathiolane ring. Nevertheless, compound **C48** has exhibited increased inhibitor effect due to hydrogen bond interaction between cyano group and Asp623 residue, when the cyano group is at the fifth position of 1,3-oxathiolane ring.

While compounds **C20-C28** which carrying chloro methyl group on the 4' carbon atom of cytidine shown that similar active site interactions to compounds **C2-C12**, when the ribose ring is replaced by the cyclopentene ring, substitution with azido and cyano group caused increased binding affinity. In addition to, although the other our designed compounds which have straight and branched chains instead of ribose, exhibit significant interactions with the active site of the target enzyme, the presence of a 5-membered cyclic ring was found to be essential for activity due to decreased binding affinity.

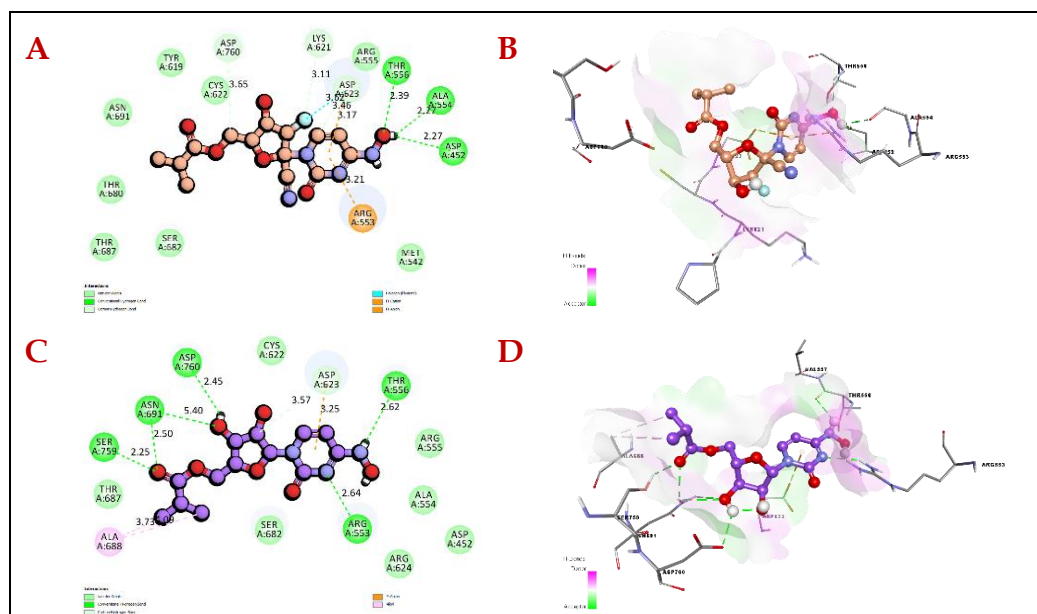


Figure 3. A) Interactions between compound **C17_PD** and SARS-CoV-2 RdRp as 2D diagram. B) Binding orientation of compound **C17_PD** in the active site of SARS-CoV-2 RdRp. c) Interactions between compound **MLN** and SARS-CoV-2 RdRp as 2D diagram. D) Binding orientation of compound **MLN** in the active site of SARS-CoV-2 RdRp.

As known, **MLN** is a prodrug and isobutyric acid ester form of **C1**. After determination of the most active compound **C17** according to the molecular docking results, isobutyric acid ester form of **C17** (**C17_PD**) was designed to evaluate the SARS-CoV-2 RdRp inhibition potential of the possible unchanged prodrug molecule. The interactions of both **MLN** and **C17_PD** with the active site are given Figure 3. In spite of hydrogen bonding interactions were detected between ester groups (C=O) of **MLN** and Asn691 and Ser759 residues, the mentioned interactions disappeared for **C17_PD** in comparison with active form **C17**. These findings are also supported by decreased binding energy.

Finally, as **MLN** undergoes triphosphorylation after hydrolysis of the ester group to show its mutagenic effect on the RNA strain, the monophosphate derivatives of both **MLN** (**MLN_MP**) and **C17** (**C17_MP**) were docked into the target active site and compared for their binding affinity (Figure 4). While the hydrogen bonding interactions of **MLN_MP** with Asn691 and Ser759 residues are continuing, monophosphate moiety of **C17_MP** interacted with Asp623 as a hydrogen bond acceptor besides Van der Waals interactions with Asp760 residue.

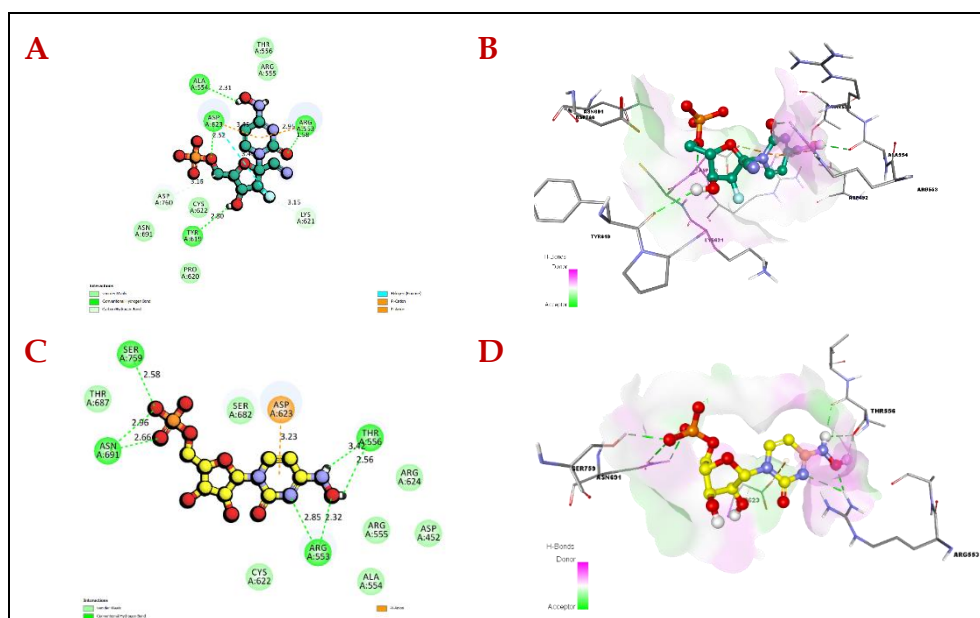


Figure 4. A) Interactions between compound **C17_MP** and SARS-CoV-2 RdRp as 2D diagram. B) Binding orientation of compound **C17_MP** in the active site of SARS-CoV-2 RdRp. c) Interactions between compound **MLN_MP** and SARS-CoV-2 RdRp as 2D diagram. D) Binding orientation of compound **MLN_MP** in the active site of SARS-CoV-2 RdRp.

The obtained docking results have shown that, compound **C17** which has been designed in this study, is the best candidate, and its prodrug **C17_PD** and the monophosphate derivative **C17_MP** exhibits more inhibitory potential than both **MLN** and compound **C1_MP**.

2.2. Analysis of bioavailability, drug-likeness and ADME/T profile

Computational methods present an advantage of being able to estimate bioavailability and safety of drug candidates and to select better lead compounds even before they are synthesized and undergo preclinical tests and clinical trials [19].

2.2.1. Analysis of bioavailability and drug-likeness

As well known, it is the main issue for effectiveness that a drug molecule must reach the target in sufficient concentration. Molecular weight (MW), number of hydrogen acceptors (nON), number of hydrogen donors (nOHN), number of rotatable bonds (nRot), topological polar surface area (TPSA), octanol-water partition coefficient (LogP_{ow}) and aqueous solubility (LogS) are main characteristics that affect membrane permeability, lipophilicity and solubility therefore bioavailability and drug-likeness of a compound. The physico-chemical, pharmacokinetic, drug-likeness and related parameters of the compounds in this study were evaluated by using the SwissADME Web tool. Table 2 lists bioavailability and drug-likeness characteristics of our compounds selected as they showed more interactions with the active site of the SARS-CoV-2 RdRp enzyme (please find Table S2 supports bioavailability and drug-likeness characteristics of all compounds in this study). MW of all compounds were found lower than 500 Da and the MW of **C17** (which is the most potent compound in the study) is 286.22 g/mol and also, MW of **C17_PD** (prodrug of **C17**) is 356.31 g/mol (MW of **MLN**:329.31 g/mol and MW of EIDD-1931, active form of **MLN**:259.22 g/mol). Other physicochemical properties (nON, nOHN, nRot, TPSA) of the **C17_PD** were found to show similarity with **MLN** as such in similarity of **C17** with EIDD-1931. All selected compounds

were estimated to have high aqueous solubility. None of the selected compounds exceed the range limit of LogP. Also, **C17** and **C17_PD** were found to have better lipophilicity (LogP) than EIDD-1931 and MLN.

There are different type rule-based approaches often originating from analyses by major pharmaceutical companies aiming to improve the quality of their proprietary chemical collections; and two of them the Lipinski (Pfizer) and the Muegge (Bayer) methods were used to evaluate the drug-likeness of the compounds in this study. Therefore, selection criteria for an ideal drug candidate molecule according to rules established by Lipinski et al. [20] are that MW < 500 Da, Log P < 5, nON ≤ 10, nOHN ≤ 5 and according to rules established by Muegge et al. [21] are that 200 ≤ MW ≤ 600, -2 ≤ Log P ≤ 5, TPSA ≤ 150 Å², number of rings ≤ 7, number of C>4, number of heteroatom > 1, nROT ≤ 15, nON ≤ 10, nOHN ≤ 5. Compounds violating more than one of these rules can pose low oral bioavailability. Only, **C13** was found to exhibit 2 violations and other compounds have been found with one or less violation. **C17_PD** and **C17** were found to have no violation.

Table 2. List of molecular properties (physico-chemical, lipophilicity, water solubility, drug likeness) of the selected compounds by using online webserver SwissADME.

Compound	Physicochemical Properties					Lipophilicity	Water solubility	Drug-Likeness	
	MW (g/mol)	nON	nOHN	nRot	TPSA (Å ²)	LogP _{o/w}	LogS (ESOL)	Lipinski Rule nviol	Muegge Rule nviol
C1 (EIDD-2801)	259.22	7	5	3	137.07	-1.82	-0.48	0	0
C13	284.23	8	5	3	160.86	-2.27	0.01	0	2
C14	268.23	7	4	3	140.63	-1.54	-0.18	0	1
C15	268.23	7	4	3	140.63	-1.56	-0.18	0	1
C17	286.22	8	4	3	140.63	-1.31	-0.31	0	1
C18	286.22	8	4	3	140.63	-1.34	-0.31	0	1
C19	304.21	9	4	3	140.63	-1.02	-0.57	0	0
C30	314.25	10	5	4	186.82	-1.68	-0.63	1	1
C32	302.22	10	4	4	166.59	-1.11	-1.10	1	1
C34	316.25	10	4	4	166.59	-0.70	-1.29	1	1
C48	270.27	6	3	3	145.70	-0.78	-0.49	0	0
MLN	329.31	8	4	6	143.14	-0.88	-0.83	0	0
C17-PD	356.31	9	3	6	146.70	-0.31	-1.17	0	0

Abbreviations: MW: Molecular weight; nON: Number of hydrogen acceptors; nOHN: Number of hydrogen donors; nRot: Number of rotatable bonds; TPSA: Topological polar surface area, LogP_{o/w}: Octanol-water partition coefficient; LogS (ESOL): Estimating aqueous solubility from molecular structure; MLN: Molnupiravir, **C17_PD**: prodrug form of compound **C17**.

2.2.2. Analysis of ADME and toxicity profile parameters

The computational models that can predict ADMET profiles have become an alternative and complementary approach to existing *in vitro* and *in vivo* toxicity tests, thereby minimizing the need for animal testing, reducing the cost and time of relevant tests, and improving bioavailability/toxicity prediction and efficacy/safety assessments [19,22]. The ADMET profile of a xenobiotic is in a close relationship with bioavailability and drug-like properties, however findings were presented in the different sections and tables to discuss easily. pkCSM [23], eMolTox, ProToxII and admetSAR, free open sources and comprehensive computer readable databases, are tools that we used to predict ADME/T-associated properties of our compounds selected due to their superior interactions with the active site of the SARS-CoV-2 RdRp enzyme. Also, SwissADME [24] and Molinspiration web tools and OSIRIS data warrior software [25] were applied to evaluate absorption and medicinal chemistry profiles (volume of molecules, drug-likeness score etc.) of the selected compounds.

Table 3. Absorption, Distribution, Metabolism and Excretion (ADME) and medicinal chemistry properties of the selected compounds.

Parameters		C13	C14	C15	C17	C17-PD	C18	C19	C30	C32	C34	C48	MLN	EIDD-1931 (C1)
Absorption	Caco2 permeability ^a	0.377	0.077	0.424	0.073	0.428	0.442	0.446	0.347	0.05	0.045	-0.007	0.498	0.425
	GI absorption ^a	low	low	low	low	low	low	low	low	low	low	low	low	low
	P-glycoprotein substrate ^b	no	no	no	no	no	no	no	no	no	no	no	no	no
	Volume ^c	227.5	219.46	219.46	224.42	294.32	224.42	229.02	251.77	232.45	248.69	212.74	280.87	210.97
	Human intestinal absorption ^a (%)	46	58.334	61.277	55.033	57.528	55.006	61.484	47.488	55.331	56.163	62.618	52.006	39.275
Bioavailability score	0.55	0.55	0.55	0.55	0.55	0.55	0.55	0.55	0.11	0.11	0.11	0.55	0.55	0.55
Distribution	VD _{ss} (human, log L/kg)	-0.525	-0.789	-0.561	-0.749	0.282	-0.472	-0.709	-0.27	-0.495	-0.476	-0.682	0.847	-0.147
	BBB permeability ^b (log BB)	-1.345	-1.248	-1.129	-1.396	-1.155	-1.245	-1.292	-1.565	-1.648	-1.67	-1.136	-1.02	-1.225
	CNS permeability ^a (log PS)	-4.186	-3.745	-3.385	-3.742	-3.335	-3.381	-3.42	-3.516	-3.451	-3.432	-3.304	-3.739	-4.124
Metabolism	CYP1A2 substrate	no	no	no	no	no	no	no	no	no	no	no	no	no
	CYP2C19 substrate	no	no	no	no	no	no	no	no	no	no	no	no	no
	CYP2C9 substrate	no	no	no	no	no	no	no	no	no	no	no	no	no
	CYP2D6 substrate	no	no	no	no	no	no	no	no	no	no	no	no	no
	CYP3A4 substrate	yes	yes	yes	yes	no	yes	yes	yes	yes	yes	yes	yes	no
	UGT catalyzed	no	no	no	no	yes	yes	yes	no	no	yes	no	yes	no
Excretion	Total Clearance ^a (log ml/min/kg)	0.603	0.607	0.612	0.524	0.088	0.512	0.601	-0.185	-0.161	-0.21	0.211	0.175	0.525
	Renal OCT2 substrate	no	no	no	no	no	no	no	no	no	no	no	no	no
Med.Chem.	PAINS ^b	0	0	0	0	0	0	0	1*	1*	1*	0	0	0
	Brenk ^b	1**	1**	1**	1**	1**	1**	1**	4***	4***	4***	1**	1**	1**
	Drug-likeness ^d	-7.48	-5.69	-8.79	-6.83	-5.8536	-6.65	-10.07	1.32	-1.97	-1.18	-4.73	-2.53	-1.89

^aThese studies were performed by using online webserver pkCSM; ^b These studies were performed by using online webserver SwissADME; ^c This parameter calculated by using Molinspiration; ^d These studies were performed by using OSIRIS data warrior software. PAINS ALERTS: *azo group BRENK ALERTS: **oxygen-nitrogen-single bond; *** azido group, diazo group, oxygen-nitrogen-single bond, quaternary nitrogen

ADME

P-glycoprotein (P-gp) that is a member of ABC transporter family, actively pumps xenobiotics out of the cells and plays an important role in the intestinal absorption and excretion of drugs and also, limits various xenobiotics enter into the central nervous system (CNS). Therefore, efficacy and safety of drug molecules related to being P-gp substrates, inhibitors or inducers [26]. The selected compounds were not predicted to be a P-gp substrate (Table 3) or inhibitor (Table 4) like MLN and active form of MLN (EIDD-1931). The human colon epithelial cancer cell line, Caco-2, is commonly used as a prediction model of human intestinal absorption and permeability of xenobiotics [27]. The selected compounds exhibited low Caco-2 permeability and human intestinal absorption (Table 3). Bioavailability scores of the selected compounds except C30, C32 and C34 were found to be similar to MLN and EIDD-1931 (Table 3).

The volume of distribution (VD_{ss}) is the theoretical value to present the distribution profile of a molecule and higher values means drug molecules distributed to tissues rather than plasma. VD_{ss} of all the selected compounds were found low and the predicted distribution profile of C17 and C17_PD shows similarity to EIDD-1931 and MLN, respectively (Table 3). The blood-brain barrier (BBB) and central nervous system (CNS) permeability of the drugs except CNS targeted drugs due to cause neurotoxicity is not expected [28]. The selected compounds have been estimated to be poorly BBB permeable and unable to penetrate the CNS (Table 3).

All selected compounds except **C17_PD** (due to isobutyric acid ester form of **C17**) were predicted to be substrates of CYP3A4 metabolism enzyme that is responsible for metabolism of many drugs (Table 3).

Total clearance refers to the combination of hepatic and renal clearance of a molecule and predicted total clearance values of the selected compounds were shown in Table 3 in log ml/min/kg. Also, compounds were tested to be a renal OCT2 substrate and none of the compounds have been expected to be a renal OCT2 substrate according to the pkCSM prediction tool.

Toxicity

The compounds selected according to their highest affinity prediction with the active site of the SARS-CoV-2 RdRp, were evaluated *in silico* for different hazard endpoints, structural alerts, disruption potential on Tox21 pathways, target organ toxicity potential and inhibition potentials to major drug metabolism enzymes and drug transporters. Table 4 lists the predicted toxicity properties of the selected compounds.

LD₅₀ refers to the degree of acute toxicity of xenobiotics and corresponds to doses that are likely to kill 50% of the animals in a batch used for experimentation. Therefore, LD₅₀ value is an important parameter to classify the toxicity of a compound. Both oral rat acute toxicity (LD₅₀, mol/kg) values (predicted by online web server pkCSM) and predicted LD₅₀ (mg/kg) values (predicted by online web server Protox-II) of the selected compounds were shown in Table 4. All compounds were classified in category 4 according to the GHS classification (Category IV; chemicals with 300 < LD₅₀ ≤ 2000 mg/kg) [29]. LOAEL refers to the low observed adverse effect level of the xenobiotics in the preclinic chronic toxicity studies and predicted LOAEL values of the selected compounds were shown in Table 4. Predicted LOAEL values of the **C17** and **C17_PD** are 2.083 log mg/kg_bw/day and 2.824 og mg/kg_bw/day, respectively.

None of the compounds is expected to exhibit cytotoxicity according to the *in silico* test results (Table 4). Predicted AMES mutagenicity test results of the selected compounds were positive, unlike **MLN** and EIDD-1931. All selected compounds, **MLN** and EIDD-1931 were predicted to have DNA damage potential (genotoxicity) and also **C17_PD** were found inactive for the carcinogenicity potential similar to **MLN** (Table 4). Structural alerts (also known as toxicophores/toxic fragments) are chemical substructures that indicate or associate to specific toxic endpoints and are widely accepted in chemical toxicology and regulatory decisions. Brenk and Pan Assay Interference Compounds (PAINS) filters in the SwissADME web tool were applied and also structural alerts in the MolTox web tool were scanned. Compounds **C30**, **C32** and **C34** were predicted to contain four Brenk alerts and one PAINS alert related to the reactivity potential of the azido group on the 4' carbon of cytidine. On the other hand, **C17** and **C17_PD** exhibited no PAINS alert and showed similarity with EIDD-1931 and **MLN** according to the Brenk filter. All selected compounds contain structural alerts for DNA damaging potential and hepatotoxicity risk like **MLN** and its active form EIDD-1931. The details of these structural alerts were shown in Table 4.

All compounds were scanned for the disruption potential on Tox21 pathways [30] and **C17** was found to active for p53 tumor suppressor gene pathway like EIDD-1931 (Table 4).

Leading to cardiotoxicity thoroughly QT prolongation related with inhibition of potassium channel encoded by ether-a-go-go-Related gene (hERG) caused withdrawals of many drugs from pharmaceutical market. So, prediction of hERG inhibition is important during drug development to prevent future failure of drug candidates [29]. Also, cardiac complications are one of the main concerns of the COVID 19 viral infections. Consequently, treatment of the disease is expected not to cause cardiotoxicity. Any compounds were not found to exhibit hERG inhibition potential, like **MLN** and EIDD-1931. None of the compounds were found to have skin sensitisation potential (Table 4). All selected compounds, **MLN** and EIDD-1931 were found related to hepatotoxicity risk. Drug-induced liver injury is one of the main concerns due to severe hepatic complications of the COVID-19 viral infection and polypharmacy in treatment [31]. Therefore, there is a need for further evaluation in detail if **MLN** and our designed compounds increase the risks of liver injury.

Inhibition of drug transporters and metabolism enzymes mainly play a role in drug interactions and drug related renal/hepatic toxicity [32,33]. All compounds were found as organic anion transporting polypeptide 2B1 (OATP2B1) and organic anion transporting polypeptide 1B1 (OATP1B1) inhibitors and these features can be increases drug interaction potential of **MLN** and our designed **MLN** analogs with the substrates of these OATP transporters. In addition, **MLN** and EIDD-1931 are cytochrome P450 3A4 (CYP3A4) inhibitors, unlike our designed compounds.

Table 4. List of predicted toxicity properties (hazard endpoints, Tox21 pathway interaction, target organ toxicity, inhibition properties) of the selected compounds.

ENDPOINT		C13	C14	C15	C17	C17- PD	C18	C19	C30	C32	C34	C48	MLN	EIDD- 1931 (C1)	
Hazard Endpoints	Oral Rat Acute Toxicity (LD 50, mol/kg) ^a	2.192	2.185	2.152	2.189	2.088	2.101	2.188	2.129	2.207	2.209	2.257	2.214	2.147	
	Predicted LD 50 (mg/kg) ^b	1000	1000	1000	1000	826	1000	1000	826	1000	1000	3320	826	826	
	Toxicity class ^b	4	4	4	4	4	4	4	4	4	4	4	4	4	
	LOAEL (log mg/kg_bw/day) ^a	2.5	1.997	1.751	2.083	2.824	2.091	1.675	2.174	1.766	1.732	1.564	3.053	2.742	
	Cytotoxicity ^b	inact.	inact.	inact.	inact.	inact.	inact.	inact.	inact.	inact.	inact.	inact.	inact.	inact.	inact.
	Genotoxicity ^c (structural alerts)	yes ¹	yes ²	yes ¹	yes ^{1,3}	yes ^{1,3}	yes ^{1,3,4}	yes ^{1,2,3,4}	yes ^{1,5}	yes ^{1,5}	yes ^{1,5}	yes ¹	yes ¹	yes ^{1,2,6}	
	Ames Toxicity (mutagenicity) ^d	yes	yes	yes	yes	yes	yes	yes	yes	yes	yes	yes	yes	no	no
	Carcinogenicity ^b	active	active	active	inact.	inact.	active	active	active	active	active	active	active	inact.	active
Immunotoxicity ^b	inact.	inact.	inact.	inact.	inact.	inact.	inact.	inact.	inact.	inact.	inact.	inact.	inact.	inact.	
Tox 21 Pathway interaction	Tox21-Nuclear receptor signaling pathways ^b	inact.	inact.	inact.	inact.	inact.	inact.	inact.	inact.	inact.	inact.	inact.	inact.	inact.	
	Tox21-Stress response pathways; -Phosphoprotein (Tumor Suppressor) p53 ^b	active	inact.	inact.	active	inact.	inact.	inact.	inact.	inact.	inact.	inact.	inact.	active	
Target Organ Toxicity	Hepatotoxicity (DILI) ^c	yes ⁷	yes ⁷	yes ^{7,8}	yes ^{7,8}	yes ⁷	yes ⁷	yes ⁷	yes ⁷	yes ⁷	yes ⁷	yes ⁸	yes ⁷	yes ^{7,9}	
	Cardio-toxicity	hERG I inhibitor ^a	no	no	no	no	no	no	no	no	no	no	no	no	no
		hERG II inhibitor ^a	no	no	no	no	no	no	no	no	no	no	no	no	no
	Skin Sensitisation ^a	no	no	no	no	no	no	no	no	no	no	no	no	no	no
Inhibition properties ^d	OATP2B1 inhibitor	no	no	no	no	no	no	no	no	no	no	no	no	no	
	OATP1B1 inhibitor	yes	yes	yes	yes	yes	yes	yes	yes	yes	yes	yes	yes	yes	
	OATP1B3 inhibitor	yes	yes	yes	yes	yes	yes	yes	yes	yes	yes	yes	yes	yes	
	MATE1 inhibitor	no	no	no	no	no	no	no	no	no	no	no	no	no	
	BSEP inhibitor	no	no	no	no	no	no	no	no	no	no	no	no	no	
	P-glycoprotein inhibitor	no	no	no	no	no	no	no	no	no	no	no	no	no	
	CYP3A4 inhibition	no	no	no	no	no	no	no	no	no	no	no	yes	yes	
	CYP2C9 inhibition	no	no	no	no	no	no	no	no	no	no	no	no	no	
	CYP2C19 inhibition	no	no	no	no	no	no	no	no	no	no	no	no	no	
	CYP2D6 inhibition	no	no	no	no	no	no	no	no	no	no	no	no	no	
CYP1A2 inhibition	no	no	no	no	no	no	no	no	no	no	no	no	no		

^aThese studies were performed by using online webserver pkCSM; ^b These studies were performed by using online webserver Protox-II; ^c These studies were performed by using online webserver eMolTox; ^d These studies were performed by using online webserver admetSAR. **STRUCTURAL ALERTS:** ¹covalent bind with DNA; ² agonist of the p53 signaling pathway; ³ potential electrophilic agents; ⁴ metabolic activation; ⁵ PAINS compounds; ⁶ Agonist of H2AX. **DILI (Drug Induced Liver Injury) ALERTS:** ⁷ Antagonist of the constitutive androstane receptor (CAR) signaling pathway; ⁸ Block OATP1B1 Transporter; ⁹ Cytotoxicity in HepG2 cells. Abbreviations: LD50: Letaal Dose 50; LOAEL: Low Observed Adverse Effect Level

Painter et al. reported the first phase I clinical trial (clinicalTrials.gov identifier: NCT04392219) results as MLN present good tolerability and dose-proportional pharmacokinetics which is not affected by food consumption following administration to healthy volunteers [34]. According to another Phase I clinical trial (clinicalTrials.gov identifier: NCT04746183) results, MLN was reported as safe and well tolerated and recommended 800 mg twice daily dose for 5 days for Phase II evaluation by Khoo et al. [34]. Any serious adverse effect was not reported by both phase I clinical trials and nowadays phase II/III clinical trials (clinicalTrials.gov identifiers: NCT045755597, NCT045755584, NCT04405570, NCT04939428, etc.) of MLN are ongoing (<https://clinicaltrials.gov>). Therefore, MLN is a promising oral antiviral agent against SARS-CoV-2,

fifty four designed analog of **MLN** were tested *in silico* for RdRp inhibition, bioavailability, ADME and safety profile and compared with **MLN**. The ADME/T profile of **C17** and **C17_PD**, which is more potent than EIDD-1931 and **MLN**, respectively for the target site of the SARS-CoV-2 RdRp enzyme were found to be similar to **MLN** and its active form EIDD-1931.

3. CONCLUSION

In this study, the fifty-four **MLN** analogues, including compound **C13** the previously reported as anti-COVID-19 agent were evaluated against SARS-CoV-2 RdRp using computer-aided methods. Our docking results supported that binding affinity of **C13** against related enzyme is higher than **MLN** like in the literature. Among the designed **MLN** analogs compound **C17** (-7.3 kcal/mol binding energy value) has been found to have the highest affinity even more than compounds **C13** and **C1**. According to the molecular docking studies selected ten compounds with the highest potential enzyme inhibition including **C17** are predicted to have a similar bioavailability and safety profile to **MLN**. Therefore, future studies (*in vitro* or *in vivo*) are necessary to confirm the antiviral activity of the **C17** by various prodrug forms.

4. MATERIALS AND METHODS

4.1. Molecular docking studies

4.1.1. Preparation on enzyme

The structure of SAR-CoV-2 RNA-dependent RNA polymerase (pdb code: 6m71) [35] was downloaded from the RCSB website with a reported resolution of 2.90 Å, and used as a target for the novel molnupiravir analogs. RNA Dependent RNA polymerase is a multimeric protein and the selected crystalized structure is completed by attachment of three additional protein peptides (nsp7-nsp8, and one additional nsp8) to the core polymerase which is chain A and contains 851 amino acids residues. In this study, we prepared the structures of RdRp without cofactors to investigate the binding affinity of the drugs. This structure without crystallized ligand was protonated using the The AutoDock Tools program and thereafter the obtained structure was energy-minimized.

4.1.2. Preparation of ligands

Besides **C1** (EIDD-1931), **MLN** and monophosphate derivatives of related compounds, the designed **MLN** analogs, compounds **C2-C55** were drawn by using the Spartan 04 software[36] (SPARTAN 04, Wavefunction, Inc., Irvine, USA) and optimized for each compound with the semi-empirical PM3 method. For each compound, the most stable conformation was selected for using in docking calculation and these pdb files converted to pdbqt files with The AutoDock Tools program [37].

4.1.3. Docking studies and analysis

AutoDock Vina software [38] was used for the designed compounds into the SARS-CoV-2 RdRp structures docking calculations. The results files were analyzed using Accelrys Discovery Studio Visualizer 4.0 program. The size of grid box was detected 30x30x30 points in *x* (114.52), *y* (114.11), and *z* (122.91) dimensions was built; appropriate to the positions of active site residues Asp760 and Asp761 in the 6m71 structure[39]. The Vina parameter "exhaustiveness" was set to the value of 10, besides a grid spacing of 0.375 Å was employed for the calculation of the energetic map of this enzyme. During the docking studies, we used flexible ligand in rigid protein. As a result of the calculation, 9 different conformations in which the ligand can bind to the enzyme were obtained, and this process was repeated three times for each ligand. For each compound, binding affinities of 27 conformations in total were obtained and selected the ten best active compounds after evaluation of these data.

4.2. Analysis of bioavailability, drug-likeness and ADMET profile

4.2.1. Analysis of bioavailability and drug-likeness

The evaluation of pharmacokinetic properties of a new drug candidate is a major step for the drug discovery and development process. SAR and QSAR models provide *in silico* rapid and accurate evaluation of the candidates and selection of the molecule that present better characteristic properties in early stages of drug development. The freely accessible servers including Swiss-ADME (<http://www.swissadme.ch/>), pkCSM (<http://biosig.unimelb.edu.au/pkcsm>) and Molinspiration (<https://www.molinspiration.com/>),

and also software called OSIRIS Datawarrior software were used to predict the molecular and ADME profile of the selected compounds that highest interacted with the active site of the SARS-CoV-2 RdRp enzyme.

4.2.2. Analysis of ADME and toxicity profiles

Computational based *in silico* toxicity measurement has been widely used due to their accuracy, rapidity, accessibility, which can provide information about any compound. To virtually identify the toxicity and adverse effect of the selected compounds that highest interacted with the active site of the SARS-CoV-2 RdRp enzyme, we used freely accessible online web servers including pkCSM, eMolTox (<https://xundrug.cn/moltox>), admetSAR 2.0 (<http://lmmd.ecust.edu.cn/admetSAR2/>) and ProTox-II (https://tox-new.charite.de/prottox_II/). Each tool was used to predict several safety parameters such as acute toxicity, cytotoxicity, carcinogenicity, mutagenicity, immunotoxicity, target organ toxicity, structural alerts related to different toxic effect mechanisms and inhibition potential of the compounds to the several metabolism enzymes and drug transporters.

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Appendix A. Supplementary Material

Supplementary material related to this article can be accessed at <https://dx.doi.org/10.29228/jrp.93>.

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