

## ESTRODİOL RESEPTÖRÜN, DOMUZ UTERUSUNDAN ELDE EDİLEN LİZOZOMCA ZENGİN FRAKSİYON TARAFINDAN İNAKTİVE EDİLMESİ

### IN VITRO INACTIVATION OF THE ESTRODIOL RECEPTOR BY A LYSOSOME-ENRICHED FRACTION FROM PIG UTERUS

Ismail PEKER\*

#### SUMMARY

Cytosol and lysosomes were prepared from pig uterus. Inactivation of estradiol receptor in cytosol was investigated with and without lysosomes from pig endometrium. Lysosomes fraction of pig uterus inactivated the binding site of estradiol receptor significantly.

**KEYWORDS** : lysosomes/estradiol receptor/pig uterus

#### ÖZET

Sitosol ve lizozom domuz uterusundan hazırlandı. Sitosol içindeki estradiol reseptörün inaktivasyonu kendi halinde veya domuz endometriumundan hazırlanan lizozom ile araştırıldı. Domuz uterusundan elde edilen lizozom estradiol reseptörün bağlanmasını önemli ölçüde inaktive etti.

#### INTRODUCTION

Research about estradiol receptor began in 1957 (1). The first specific steroidbinding protein in the cell was found by Jensen et al (2, 3). This protein was extracted with buffer and characterized by sucrogradient zentrifugation and named as receptor. In 1966 Jungblut extracted an estradiol receptor from the nucleus fraction of uteri. After injection estradiol solution, takes uterus lumen very quickly. In a few minutes an accumulation of receptor and steroid flow in the cell nuclei (4). In this time cytosol receptor is reduced. After 5-7 hours cytosol receptor concentration

\* Dr. Pakize İ. Tarzi Biochemistry and Hormon Laboratory, Valikonağı Cad. No. 86 - 80200 Nişantaşı - İSTANBUL.

increases again. However in the cell nuclei estradiol receptor sinks during this time. Nearly 40 % of estradiol receptors come again in the cell nuclei, but the rest 60 % is synthesized new in the cytoplasm. It has been shown that estradiol modifies the level of its receptor in both the rat (Jensen et al 1969 (5), Gorski et al 1970 (6), and the pig uterus (Jungblut et al 1976 (7) by enhancing receptor biosynthesis. In vitro studies by Coulson and Pavlik 1977 have indicated that cytosol is not involved in receptor degradation, while Peck et al (1973) (8), suggested that the instability of the estrogen receptor might result from its susceptibility to lysosomal attack. How does estradiol receptor metabolize, and is there for it or is it the work of the protease enzyme? Do lysosomal enzymes play a role in the inactivation of estradiol receptor? We isolated lysosomes from pig uterus and incubated estradiol receptor in cytosol. We investigated the inactivation of estradiol receptor with and without lysosomal enzymes.

## MATERIAL AND METHODS

The preparation of cytosol, electrophoresis were carried out to Hekim, N and Jungblut P.W (9). The preparation of mitochondria-lysosomes and lysosomes fraction and determination of enzymes were done according to Sierralta, W et al (10).

Radioactive estradiol came from Amersham, agar purum and the protein determination kit from Behringwerke, the rest of chemicals were purchased from merck.

Measurement of radioactivity : Samples of extracts and fraction of density gradient analysis were counted in polyethylene vials after addition of 15 ml of scintillation fluid (80 g naphthalene, 5 g PPO, 50 mg POPOP/1000 ml xylol-dioxane 1:2) in Packard Tri-Carbs model 3320 with tritium efficiencies of 40-46%.

## RESULTS AND DISCUSSION

Isolation of lysosomes from pig uterus : The pig uterus homogenate in sucrose gradient was centrifuged and fractionated. We used N-acetyl  $\beta$ -glucosaminidase for lysosome and ICDH for mitochondria as marker enzyme which were measured in each fraction. The peaks collected as lysosome, and mitochondria-lysosomes fraction respectively.

Lysosomes was separated from pig uterus and tested as flowing.

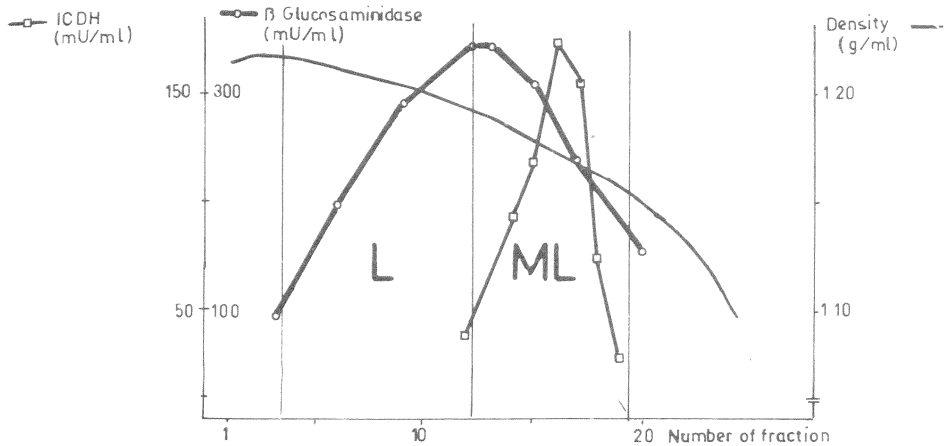


Fig - 1 : Isopycnic subfraction of a mitochondrial/lysosomal fraction from pig endometrium was homogenized with buffered sucrose and separated on a 25-52 %, w/w sucrogradienten. 2 ml of a 1:6 suspension of a 17500 g pellet from endometriumhogenat, in 50 mM tris, 3mM EDTA, 0.25 M sucrose pH 7.5 was carried out on a 27 ml of a 25-52 % in sucrogradienten. After 15 hours centrifugation at 10 000 rpm 1 °C the activity of enzymes were measured in the fraction and then pooled. The density was calculated with refraktion index.

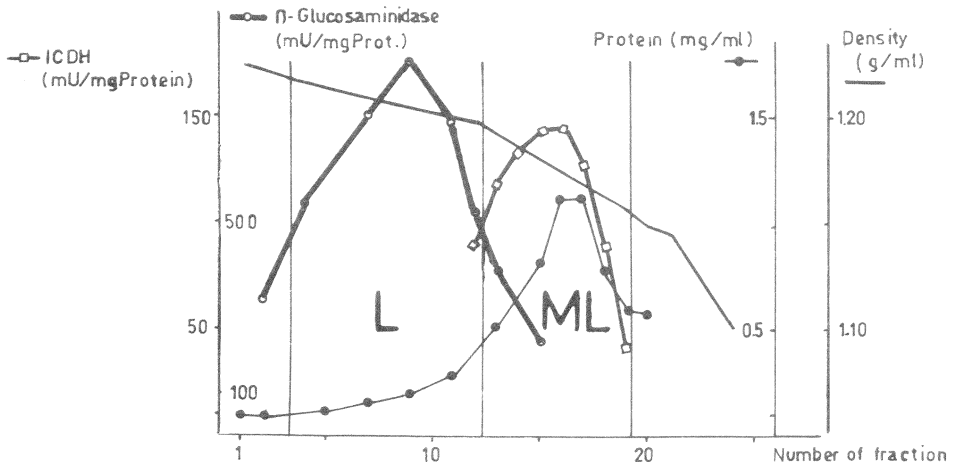


Fig - 2 : The specific activity of  $\beta$ - glucosaminidase and ICDH of a ML and L. The Specific activity of fraction was calculated enzymactivity and protein concentration.

**Table - 1 :** HM : Homegenat, ML : Mitochondria-Lysosomes, L : Lysosomes

Protein concentration (mg/dl)		N-acetyl- $\beta$ -glukose-aminidase (mu/ml) (mu/mg prot)		ICDH (mu/ml) (mu/mg prot)		PURIFICATIO fold
HM	45	1890	42	1350	30	1
ML	14	2590	185	1446	103	4.4
L	0.72	610	847	-	-	20

2 pellets of L containing 0.72 mg protein were dissolved in 250  $\mu$ l of the buffer (10 mM Tris + 4mM EDTA + 5 mM DTT + 0.5 % surfynol) and incubated for 24 hr, at 4 °C, then frozen and thawed three times. After then cytosol was incubated with this lysosomes suspension or mitochondria-lysosomes fraction from pig uterus endometrium.

**Table - 2 :** These mixtures (I, II, III) were incubated at room temperature for 1 hr, then in a cold room for 24 hr, then were done electro-phoresis. Agargel was cut in pieces. The pieces were measured in scintilation liquid in the counter.

	I	II	III
Cytosol	100 $\mu$ l	100 $\mu$ l	—
Lysosomes sus	100 $\mu$ l	—	100 $\mu$ l
E2 ( $2.10^{-7}$ )	12.5 $\mu$ l	12.5 $\mu$ l	12.5 $\mu$ l
Buffer	37.5 $\mu$ l	137.5 $\mu$ l	137.5 $\mu$ l
total	250 $\mu$ l	250 $\mu$ l	250 $\mu$ l

**Table - 3 :** Inactivation of estrodiol receptor in cytosol with lysosomes fraction.

	(+) site of gel	(-) site of gel	total
Cytosol	2736 cpm	2358 cpm	5904 cpm
cyt+lys	564 cpm	2076 cpm	2640 cpm
Lysosomes	202 cpm	882 cpm	1084 cpm
Inact.	79.4 %	12.9 %	48.2 %

### CONCLUSION

The effect of lysosomes enzymes on estrodiol receptor is more at (+) site than (-) site of the agar gelectrophoresis. CPM in lysosomes may occur during the contamination of estrodiol receptor with lysosome-enriched fraction but may however come from the background of counter.

### REFERENCES

1. Jensen, E.V., Jacobson, H.I. : *Recent Prog. Horm. Res.*, **18**, 387 (1962).
2. Jensen, *et all* : In *Steroid Dynamics*, Fincus, G., Nakao, I. and Tait, J.F., Edit by, Academic Press, N.Y., p. 133, (1966).
3. Jensen, E.V., Jacobson, H. I. : Pincus, G. and Vollmer, E.P., Ed. Academic Pres, N.Y., p. 161, (1960).
4. Jungblut, P.W. *et all* : *J. Steroid Biochem.*, **11**, 273-278 (1979).
5. Jensen E.V., Suzuki, T., Numata, M., Smith, S., Desombre, ER : *Steroids*, **13**, 417-427 (1969).
6. Gorski, J., Sarff, M., Clark, J.H., (1970) : In Raspe G (Ed), *Adv. in the Biosciences* 7. edn, Vieweg, Braunschweig, pp. 1-20, (1970).
7. Jungblut, P.W., Gaus, J., Hughes, A., Kallweit, E., Sierralta, W., Szendro, P., Wagner, R. : *J. Steroid Biochem.*, **7**, 1109-1116 (1976).
8. Peck *et all* : *Biochemistry*, **12**, 4603-4608 (1973).
9. Hekim, N., Jungblut, P.W. : *Hoppe-Seylers Z. Physiol. Chem.*, **364**, 607-611 (1983).
10. Maria, A., Sierralta, W. : *Acta Endocrinologica*, **98**, 295-301 (1981).

(Received January 12, 1992)