

Design, synthesis and *in vitro* evaluation of new thiosemicarbazone derivatives as potential anticancer agents

Belgin SEVER ¹, Gülşen AKALIN ÇİFTÇİ ², Ahmet ÖZDEMİR ¹, Mehlika Dilek ALTINTOP ^{1*}

- ¹ Department of Pharmaceutical Chemistry, Faculty of Pharmacy, Anadolu University, 26470 Eskişehir, Turkey.
- Department of Biochemistry, Faculty of Pharmacy, Anadolu University, 26470 Eskişehir, Turkey.
- * Corresponding Author. E-mail: mdaltintop@anadolu.edu.tr (M.D.A); Tel. +90-222-3350580/3772; Fax: +90-222-3350750. ORCID No: 0000-0002-8159-663X.

Received: 01 June 2018 / Revised: 17 August 2018 / Accepted: 31 August 2018

ABSTRACT: Thiosemicarbazones play a pivotal role as potential therapeutic agents for the management of lung cancer. 4-(4-Cyanophenyl)-1-[(5-arylfuran-2-yl)methylene]thiosemicarbazides (1-10) were obtained *via* the reaction of 4-(4-cyanophenyl)thiosemicarbazide with 5-arylfurfurals. MTT assay was performed to assess their cytotoxic effects on A549 human lung adenocarcinoma and L929 mouse fibroblast cell lines. Compounds **1**, **5**, **6** and **7** were identified as the most effective anticancer agents on A549 cell line with IC50 values of $12.75\pm0.35~\mu g/mL$, $4.30\pm0.61~\mu g/mL$, $5.50\pm2.12~\mu g/mL$ and $5.90\pm0.57~\mu g/mL$, respectively compared to cisplatin (IC50 = $12.00\pm0.71~\mu g/mL$). The IC50 values of these compounds for L929 cell line were higher than their IC50 values for A549 cell line indicating that their anticancer effects were selective. The apoptotic effects of these compounds were also analyzed based on Annexin V-PI binding capacities in flow cytometry. According to flow cytometric analyses, the early apoptotic effects of compounds **1**, **5**, **6** and **7** on A549 cell line were determined as 4.7, 5.1, 7.3 and 5.1%, whereas their late apoptotic effects were determined as 3.7, 2.0, 4.9 and 3.3%, respectively. Compound **6** showed more apoptotic activity than compounds **1**, **5** and **7**. According to these findings, compounds **5** and **6** stand out as promising anticancer agents for further *in vitro* and *in vivo* studies.

KEYWORDS: Apoptosis; cytotoxicity; furan; lung cancer; thiosemicarbazone.

1. INTRODUCTION

Lung cancer remains the leading cause of cancer-related death across the globe with a 5-year survival rate of less than 15% [1,2]. As the most common lung cancer, non-small cell lung cancer (NSCLC) poses a major threat to public health. Despite substantial advances in diagnostic and therapeutic approaches, the overall survival for NSCLC patients still remains poor [3]. One main impediment for the treatment of NSCLC is that most patients are diagnosed at advanced or metastatic stages (stage III/IV) when the prognosis is poor and therapeutic options are limited [2-4].

Apoptosis is a tightly regulated cellular process that is essential for removal of damaged or unwanted cells and maintenance of cellular homeostasis [5,6]. In pathological conditions, particularly cancer, cells lose their ability to undergo apoptosis leading to uncontrolled proliferation [7]. Mounting evidence has shown that most of the anticancer agents trigger the induction of apoptosis and related cell death networks to eliminate tumor cells [7] and therefore the discovery of new apoptosis-inducing antitumor agents [8] is of great importance for the management of many types of cancer including lung cancer [6–10].

Thiosemicarbazones (TSCs) have attracted a great deal of attention for many years owing to their pharmacological utility as therapeutic agents and versatility as ligands allowing them to give rise to a great variety of coordination modes [11,12]. In particular, TSCs have been identified as promising antiproliferative agents against a variety of tumors through the inhibition of diverse molecular targets including ribonucleotide reductase (RNR) [11–20]. Among them, triapine (3-aminopyridine-2-carboxaldehyde thiosemicarbazone, 3-AP), a potent RNR inhibitor, is an anticancer agent currently in clinical trials (Figure 1) [11–14].

How to cite this article: Sever B, Akalın Çiftçi G, Özdemir A, Altıntop MD. Design, synthesis and *in vitro* evaluation of new thiosemicarbazone derivatives as potential anticancer agents. J Res Pharm. 2019; 23(1): 16-24.

Figure 1. Triapine.

On the basis of the aforementioned findings and in the continuation of our ongoing research efforts to identify new thiosemicarbazone derivatives as anticancer agents [18-20], herein we reported the preparation of a new series of thiosemicarbazones and focused on their cytotoxic effects on A549 human lung adenocarcinoma and L929 mouse fibroblast cell lines. The most active compounds were also evaluated for their apoptotic effects on A549 cell line.

2. RESULTS AND DISCUSSION

The synthesis of thiosemicarbazone derivatives (**1-10**) was carried out according to the steps depicted in Figure 2. In the initial step, 4-(4-cyanophenyl)thiosemicarbazide was synthesized *via* the reaction of 4-cyanophenyl isothiocyanate with hydrazine hydrate. The reaction of 4-(4-cyanophenyl)thiosemicarbazide with 5-arylfurfurals afforded 4-(4-cyanophenyl)-1-[(5-arylfuran-2-yl)methylene]thiosemicarbazides (**1-10**). The IR, ¹H NMR, ¹³C NMR, and HRMS data were in agreement with the proposed structures of compounds **1-10**.

1-10

Compound	R
1	2-C1
2	3-C1
3	4-C1
4	$2-NO_2$
5	$3-NO_2$
6	$4-NO_2$
7	2,4-Cl ₂
8	3,4-Cl ₂
9	2,5-Cl ₂
10	2-NO ₂ -4-Cl

Figure 2. The synthetic route for the preparation of the thiosemicarbazone derivatives (1-10). Reagents and conditions: (i) NH₂NH₂.H₂O, ethanol, rt, 4h; (ii) ArCHO, ethanol, reflux, 8h.

MTT assay was performed to determine the antiproliferative effects of compounds **1-10** on A549 human lung adenocarcinoma cell line. Compounds **5**, **6** and **7** were more effective on A549 cell line than cisplatin, whereas compound **1** showed similar anticancer activity against A549 cell line compared to cisplatin (IC₅₀= $12.00\pm0.71~\mu g/mL$). IC₅₀ Values of compounds **1**, **5**, **6** and **7** for A549 cell line were found as $12.75\pm0.35~\mu g/mL$, $4.30\pm0.61~\mu g/mL$, $5.50\pm2.12~\mu g/mL$ and $5.90\pm0.57~\mu g/mL$, respectively. This outcome pointed out the importance of the position of chloro and nitro substituents for anticancer activity against A549 cell line. In particular, 2-chloro, 3-nitro, 4-nitro and 2,4-dichloro substitutions significantly enhanced anticancer activity, whereas other substitutions significantly decreased anticancer activity.

In order to determine whether the compounds were toxic or nontoxic to healthy cells, the effects of compounds **1-10** on L929 mouse fibroblast cells were investigated using MTT test. The selectivity index (SI) values of compounds **1**, **5**, **6** and **7** were also determined to compare their selectivity (Table 1). The IC $_{50}$ values of these compounds for L929 cell line were higher than their IC $_{50}$ values for A549 cell line. This outcome indicated that their anticancer effects were selective. In particular, compounds **5** and **6** showed more selective anticancer activity than other compounds.

Table 1. IC₅₀ values of the compounds against A549 and L929 cells after 24 h.

Compound —	IC ₅₀ (μ ₂	IC ₅₀ (μg/mL)	
	A549 Cell line	L929 Cell line	SI*
1	12.75±0.35	21.00±1.41	1.65
2	>500	39.00±3.61	ND
3	>500	26.00±1.73	ND
4	>500	20.33±1.53	ND
5	4.30±0.61	20.00±5.00	4.65
6	5.50±2.12	23.33±1.53	4.24
7	5.90±0.57	9.67±2.08	1.64
8	>500	14.00±3.46	ND
9	>500	6.90±0.36	ND
10	>500	9.00±1.00	ND
Cisplatin	12.00±0.71	ND	ND

ND: Not Determined.

After 24 h incubation, the apoptotic effects of compounds 1, 5, 6, 7 and cisplatin on A549 cells were analyzed based on Annexin V-Propidium iodide (PI) binding capacities in flow cytometry as depicted in Figure 3. The early apoptotic effects of compounds 1, 5, 6, 7 and cisplatin on A549 cell line (at IC₅₀ values) were determined as 4.7, 5.1, 7.3, 5.1 and 9.8% respectively, whilst their late apoptotic effects were determined as 3.7, 2.0, 4.9, 3.3 and 3.5%, respectively (Table 2 and Figure 3). According to these findings, compound 6 showed more apoptotic (early and late) activity than compounds 1, 5 and 7. The results indicated that p-nitro substituent significantly enhanced apoptotic activity against A549 cell line.

Table 2. Percents of typical quadrant analysis of Annexin V FITC/PI flow cytometry of A549 cells treated with compounds **1**, **5**, **6**, **7** and cisplatin.

Groups	Early apoptotic cells%	Late apoptotic cells%	Viability%	Necrosis%
Control	1.4	0.6	96.4	1.6
Cells treated with compound 1	4.7	3.7	86.8	2.1
Cells treated with compound 5	5.1	2.0	91.1	1.8
Cells treated with compound 6	7.3	4.9	85.8	2.0
Cells treated with compound 7	5.1	3.3	89.9	1.7
Cells treated with cisplatin	9.8	3.5	85.2	1.5

A549 cells were cultured for 24 hours in medium with compounds 1, 5, 6, 7 and cisplatin at IC₅₀ values. At least 10,000 cells were analyzed per sample, and quadrant analysis was performed.

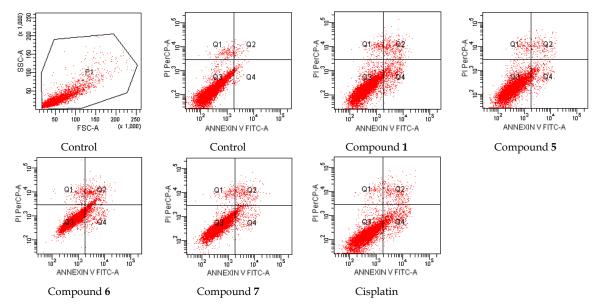


Figure 3. Flow cytometric analysis of A549 cells treated with IC_{50} values of compounds **1**, **5**, **6**, **7** and cisplatin. At least 10,000 cells were analyzed per sample, and quadrant analysis was performed.

^{*}SI= IC_{50} for L929 cell line / IC_{50} for A549 cell line.

3. CONCLUSION

In the current work, we described the synthesis of new thiosemicarbazone derivatives, which were investigated for their cytotoxic and apoptotic effects on A549 cell line. Compounds **5** and **6** were identified as the most potent anticancer agents in this series against A549 cell line with IC $_{50}$ values of 4.3±0.61 μ g/mL and 5.5±2.12 μ g/mL, respectively. Compound **6** also induced apoptosis significantly when compared with other compounds. Further studies are needed to enlighten the exact mechanism of action for the evaluation of compounds **5** and **6** as therapeutic agents for the treatment of NSCLC.

4. MATERIALS AND METHODS

4.1. Chemistry

All reagents purchased from commercial suppliers were used without further purification. The MP90 digital melting point apparatus (Mettler Toledo, Ohio, USA) was used to determine the melting points (M.p.) of the compounds. IR spectra were recorded on an IRPrestige-21 Fourier Transform Infrared spectrophotometer (Shimadzu, Tokyo, Japan). ¹H NMR and ¹³C NMR spectra were recorded on a Varian Mercury-400 FT-NMR spectrometer (Agilent, Palo Alto, CA, USA). HRMS spectra were recorded on a Shimadzu LCMS-IT-TOF system (Shimadzu, Kyoto, Japan).

4.1.1. General procedure for the synthesis of the compounds

4-(4-Cyanophenyl)thiosemicarbazide [21]

A mixture of 4-cyanophenyl isothiocyanate (0.1 mol) and hydrazine hydrate (0.2 mol) in ethanol (30 mL) was stirred at room temperature for 4 h and then filtered. The residue was crystallized from ethanol.

4-(4-Cyanophenyl)-1-[(5-arylfuran-2-yl)methylene]thiosemicarbazides (1-10)

A mixture of 4-(4-cyanophenyl)thiosemicarbazide (0.01 mol) and 5-arylfurfural (0.01 mol) was refluxed in ethanol for 8 h, filtered and crystallized from ethanol.

4-(4-Cyanophenyl)-1-[(5-(2-chlorophenyl)furan-2-yl)methylene]thiosemicarbazide (1)

M.p. 192.8 °C. Yield 80%.

IR v_{max} (cm⁻¹): 3257.77 (N-H stretching), 3138.18 (Aromatic C-H stretching), 2970.38 (Aliphatic C-H stretching), 2222.00 (C \equiv N stretching), 1602.85, 1579.70, 1537.27, 1519.91, 1492.90, 1467.83 (N-H bending, C \equiv N and C \equiv C stretching), 1411.89 (C-H bending), 1319.31, 1271.09, 1220.94, 1201.65, 1176.58, 1097.50, 1083.99, 1024.20 (C-N, C-O stretching and aromatic C-H in plane bending), 921.97, 910.40, 840.96, 800.96, 775.38, 756.10, 729.09 (Aromatic C-H out of plane bending).

 1 H NMR (400 MHz, DMSO- 2 d₆) δ (ppm): 7.30 (d, J = 3.60 Hz, 1H, aromatic proton), 7.34 (d, J = 3.60 Hz, 1H, aromatic proton), 7.37-7.41 (m, 1H, aromatic proton), 7.47-7.50 (m, 1H, aromatic proton), 7.58 (m, 1H, aromatic proton), 7.76 (d, J = 7.60 Hz, 1H, aromatic proton), 7.85 (d, J = 8.80 Hz, 2H, aromatic protons), 7.99 (d, J = 8.80 Hz, 2H, aromatic protons), 8.17 (s, 1H, -CH=N), 10.25 (s, 1H, S=C-NH), 12.22 (s, 1H, C=N-NH).

¹³C NMR (100 MHz, DMSO- d_6) δ (ppm): 102.44 (C), 106.72 (CH), 113.46 (CH), 115.64 (CH), 116.95 (C), 118.91 (C), 124.70 (2CH), 127.62 (CH, d, J= 6.5 Hz), 128.32 (CH), 129.45 (CH, d, J= 20.5 Hz), 130.83 (CH), 132.01 (2CH), 133.27 (CH, d, J= 40.4 Hz), 143.36 and 144.60 (CH), 148.94 (C), 151.01 (C), 175.13 (C).

HRMS (ESI) (m/z): [M+H]⁺ calcd. for C₁₉H₁₃ClN₄OS: 381.0571, found: 381.0582.

 $4-(4-Cyanophenyl)-1-[(5-(3-chlorophenyl)furan-2-yl)methylene] thiosemicar bazide~{\bf (2)}$

M.p. 176.6 °C. Yield 82%.

IR v_{max} (cm⁻¹): 3346.50, 3278.99 (N-H stretching), 3126.61 (Aromatic C-H stretching), 2972.31 (Aliphatic C-H stretching), 2220.07 (C \equiv N stretching), 1635.64, 1606.70, 1581.63, 1541.12, 1527.62, 1508.33, 1456.26 (N-H bending, C \equiv N and C \equiv C stretching), 1417.68 (C-H bending), 1321.24, 1273.02, 1251.80, 1201.65, 1176.58, 1099.43, 1078.21, 1060.85, 1028.06 (C-N, C-O stretching and aromatic C-H in plane bending), 997.20, 981.77, 962.48, 933.55, 871.82, 831.32, 773.46, 756.10, 684.73, 671.23 (Aromatic C-H out of plane bending).

 1 H NMR (400 MHz, DMSO- d_{6}) δ (ppm): 7.23 (d, J = 4.00 Hz, 1H, aromatic proton), 7.29 (d, J = 3.60 Hz, 1H, aromatic proton), 7.37-7.41 (m, 1H, aromatic proton), 7.46-7.51 (m, 1H, aromatic proton), 7.71-7.80 (m, 1H,

Research Article

aromatic proton), 7.83 (d, J = 8.80 Hz, 2H, aromatic protons), 7.90 (t, J = 3.60, 2.00, 1.60 Hz, 1H, aromatic proton), 7.98 (d, J = 8.80 Hz, 2H, aromatic protons), 8.12 (s, 1H, -CH=N), 10.24 (s, 1H, S=C-NH), 12.18 (s, 1H, C=N-NH).

 13 C NMR (100 MHz, DMSO- d_6) δ (ppm): 102.45 (C), 106.72 (CH), 109.92 (CH), 116.66 (CH, d, J= 58.4 Hz), 118.93 (C), 122.56 (C), 123.57 (CH), 124.78 (2CH), 127.98 (CH), 131.11 (CH, d, J= 46.1 Hz), 132.30 (2CH), 132.69 (CH, d, J= 32.7 Hz), 133.69 (CH, d, J= 39.7 Hz), 143.41 and 144.61 (CH), 149.27 (C), 153.37 (C), 175.10 (C).

HRMS (ESI) (m/z): [M+H]+ calcd. for C₁₉H₁₃ClN₄OS: 381.0571, found: 381.0564.

4-(4-Cyanophenyl)-1-[(5-(4-chlorophenyl)furan-2-yl)methylene]thiosemicarbazide (3)

M.p. 195.1 °C. Yield 85%.

IR v_{max} (cm⁻¹): 3363.86, 3288.63 (N-H stretching), 3070.68 (Aromatic C-H stretching), 2970.38 (Aliphatic C-H stretching), 2218.14 (C=N stretching), 1600.92, 1585.49, 1521.84, 1506.41, 1475.54 (N-H bending, C=N and C=C stretching), 1444.68, 1413.82, 1398.39 (C-H bending), 1321.24, 1253.73, 1209.37, 1174.65, 1093.64, 1024.20 (C-N, C-O stretching and aromatic C-H in plane bending), 979.84, 921.97, 827.46, 802.39, 783.10, 731.02, 680.87 (Aromatic C-H out of plane bending).

¹H NMR (400 MHz, DMSO- d_6) δ (ppm): 7.15-7.24 (m, 3H, aromatic protons), 7.50-7.56 (m, 2H, aromatic protons), 7.72-7.89 (m, 3H, aromatic protons), 7.99 (d, J = 8.40 Hz, 2H, aromatic protons), 8.13 (s, 1H, -CH=N), 10.22 (s, 1H, S=C-NH), 12.17 (s, 1H, C=N-NH).

¹³C NMR (100 MHz, DMSO- d_6) δ (ppm): 102.37 (C), 106.62 (CH), 109.17 (CH), 116.65 (CH, d, J= 46.8 Hz), 118.86 (C), 119.29 (C), 124.67 (2CH), 125.64 (CH, d, J= 6.4 Hz), 128.22 (CH, d, J= 9.6 Hz), 128.94 (CH, d, J= 4.5 Hz), 132.23 (2CH), 132.62 (CH, d, J= 10.3 Hz), 133.25 (CH, d, J= 34.6 Hz), 143.34 and 144.54 (CH), 148.47 (C, d, J= 11.5 Hz), 153.63 (C, d, J= 32.7 Hz), 174.97 (C).

HRMS (ESI) (m/z): [M+H]⁺ calcd. for C₁₉H₁₃ClN₄OS: 381.0571, found: 381.0590.

4-(4-Cyanophenyl)-1-[(5-(2-nitrophenyl)furan-2-yl)methylene]thiosemicarbazide (4)

M.p. 201.1 °C. Yield 86%.

IR v_{max} (cm⁻¹): 3282.84 (N-H stretching), 3118.90, 3066.82 (Aromatic C-H stretching), 2970.38 (Aliphatic C-H stretching), 2220.07 (C \equiv N stretching), 1604.77, 1516.05 (N-H bending and C \equiv N stretching), 1438.90, 1411.89 (C-H bending), 1344.38, 1325.10, 1269.16, 1255.66, 1207.44, 1176.58, 1026.13 (C-N, C-O stretching and aromatic C-H in plane bending), 981.77, 925.83, 835.18, 781.17, 744.52, 702.09 (Aromatic C-H out of plane bending).

¹H NMR (400 MHz, DMSO- d_6) δ (ppm): 7.14 (d, J = 3.60 Hz, 1H, aromatic proton), 7.27 (d, J = 3.20 Hz, 1H, aromatic proton), 7.60-7.65 (m, 1H, aromatic proton), 7.73-7.80 (m, 2H, aromatic protons), 7.86 (d, J = 8.40 Hz, 2H, aromatic protons), 7.92-7.98 (m, 1H, aromatic proton), 8.05 (d, J = 8.80 Hz, 2H, aromatic protons), 8.08 (s, 1H, -CH=N), 10.09 (s, 1H, S=C-NH), 12.29 (s, 1H, C=N-NH).

 13 C NMR (100 MHz, DMSO- d_6) δ (ppm): 102.40 (C), 106.65 (CH), 112.18 (CH), 115.39 (CH), 116.91 (C), 118.82 (C), 121.69 (CH), 123.80 (2CH, d, J= 10.3 Hz), 128.67 (CH), 129.69 (CH), 132.39 (2CH, d, J= 11.6 Hz), 133.43 (CH), 142.99 and 144.56 (CH), 146.89 (CH), 149.49 (C), 150.14 (C), 174.78 (C).

HRMS (ESI) (m/z): $[M+H]^+$ calcd. for $C_{19}H_{13}N_5O_3S$: 392.0812, found: 392.0791.

4-(4-Cyanophenyl)-1-[(5-(3-nitrophenyl)furan-2-yl)methylene]thiosemicarbazide (5)

M.p. 211.3 °C. Yield 88%.

IR v_{max} (cm⁻¹): 3321.42, 3261.63 (N-H stretching), 3163.26, 3068.75 (Aromatic C-H stretching), 2953.02 (Aliphatic C-H stretching), 2223.92 (C \equiv N stretching), 1589.34, 1517.98, 1494.83 (N-H bending, C=N and C=C stretching), 1440.83 (C-H bending), 1346.31, 1323.17, 1265.30, 1253.73, 1190.08, 1174.65, 1026.13 (C-N, C-O stretching and aromatic C-H in plane bending), 979.84, 947.05, 923.90, 896.90, 862.18, 837.11, 798.53, 781.17, 738.74, 731.02, 680.87 (Aromatic C-H out of plane bending).

 1 H NMR (400 MHz, DMSO- d_6) δ (ppm): 7.28 (d, J = 3.60 Hz, 1H, aromatic proton), 7.45 (d, J = 3.60 Hz, 1H, aromatic proton), 7.72-7.76 (m, 2H, aromatic protons), 7.83 (d, J = 8.40 Hz, 2H, aromatic protons), 7.99 (d, J = 8.40 Hz, 2H, aromatic protons), 8.14 (s, 1H, -CH=N), 8.24-8.27 (m, 1H, aromatic proton), 8.55-8.56 (m, 1H, aromatic proton), 10.25 (s, 1H, S=C-NH), 12.20 (s, 1H, C=N-NH).

 13 C NMR (100 MHz, DMSO- d_6) δ (ppm): 102.42 (C), 106.69 (CH), 110.85 (CH), 116.06 (CH), 116.92 (C), 118.19 and 118.88 (CH), 122.48 (CH), 124.55 (2CH), 129.95 (CH), 130.61 and 130.85 (CH), 132.29 (2CH), 132.90 (CH), 143.35 (CH), 148.44 (C), 149.81 (C), 152.50 (C), 175.11 (C).

HRMS (ESI) (m/z): $[M+H]^+$ calcd. for $C_{19}H_{13}N_5O_3S$: 392.0812, found: 392.0807.

4-(4-Cyanophenyl)-1-[(5-(4-nitrophenyl)furan-2-yl)methylene]thiosemicarbazide (6)

M.p. 217.4 °C. Yield 92%.

IR v_{max} (cm⁻¹): 3313.71 (N-H stretching), 3122.75 (Aromatic C-H stretching), 2976.16 (Aliphatic C-H stretching), 2223.92 (C \equiv N stretching), 1597.06, 1548.84, 1508.33, 1492.90 (N-H bending, NO₂, C \equiv N and C \equiv C stretching), 1446.61, 1411.89 (C-H bending), 1323.17, 1298.09, 1274.95, 1215.15, 1174.65, 1107.14, 1020.34 (C-N, C-O stretching and aromatic C-H in plane bending), 925.83, 848.68, 833.25, 790.81, 777.31, 752.24, 738.74, 692.44 (Aromatic C-H out of plane bending).

 1 H NMR (400 MHz, DMSO- d_6) δ (ppm): 7.32 (d, J = 3.60 Hz, 1H, aromatic proton), 7.49 (d, J = 3.60 Hz, 1H, aromatic proton), 7.74-7.79 (m, 1H, aromatic proton), 7.86 (d, J = 8.40 Hz, 2H, aromatic protons), 8.01 (d, J = 8.80 Hz, 2H, aromatic protons), 8.05-8.09 (m, 1H, aromatic proton), 8.16 (s, 1H, -CH=N), 8.29 (d, J = 9.20 Hz, 2H, aromatic protons), 10.26 (s, 1H, S=C-NH), 12.27 (s, 1H, C=N-NH).

 13 C NMR (100 MHz, DMSO- 4 6) δ (ppm): 102.39 (C), 106.75 (CH), 112.55 (CH), 116.17 (CH), 116.90 (C), 118.86 (CH), 124.34 (CH), 124.62 (2CH, d, 2 7.7 Hz), 132.28 (2CH), 132.69 (CH), 133.40 (C), 135.02 (CH), 143.27 (CH), 146.24 (CH), 150.67 (C), 152.58 (C), 175.12 (C).

HRMS (ESI) (m/z): $[M+H]^+$ calcd. for $C_{19}H_{13}N_5O_3S$: 392.0812, found: 392.0815.

4-(4-Cyanophenyl)-1-[(5-(2,4-dichlorophenyl)furan-2-yl)methylene]thiosemicarbazide (7)

M.p. 197.9 °C. Yield 84%.

IR v_{max} (cm⁻¹): 3332.99, 3292.49 (N-H stretching), 3066.82 (Aromatic C-H stretching), 2214.28 (C \equiv N stretching), 1606.70, 1587.42, 1539.20, 1477.47, 1462.04 (N-H bending, C=N and C=C stretching), 1384.89 (C-H bending), 1323.17, 1290.38, 1265.30, 1176.58, 1099.43, 1031.92 (C-N, C-O stretching and aromatic C-H in plane bending), 987.55, 921.97, 869.90, 833.25, 813.96, 786.96, 686.66 (Aromatic C-H out of plane bending).

 1 H NMR (400 MHz, DMSO- 2 d₆) δ (ppm): 7.28 (d, J = 3.60 Hz, 1H, aromatic proton), 7.34 (d, J = 3.60 Hz, 1H, aromatic proton), 7.52-7.54 (m, 1H, aromatic proton), 7.70-7.73 (m, 1H, aromatic proton), 7.82 (d, J = 9.20 Hz, 2H, aromatic protons), 7.97-8.01 (m, 3H, aromatic protons), 8.14 (s, 1H, -CH=N), 10.19 (s, 1H, S=C-NH), 12.17 (s, 1H, C=N-NH).

¹³C NMR (100 MHz, DMSO- d_6) δ (ppm): 102.41 (C), 106.68 (CH), 113.76 (CH), 115.45 (CH), 116.89 (C), 118.78 (C), 124.51 (2CH), 126.54 (C), 127.79 (CH), 129.30 (CH), 130.09 (CH, d, J= 10.3 Hz), 132.22 (2CH), 132.09 (CH, d, J= 52.6 Hz), 143.25 and 144.57 (CH), 149.17 (C), 149.98 (C), 175.14 (C).

HRMS (ESI) (m/z): [M+H]⁺ calcd. for C₁₉H₁₂Cl₂N₄OS: 415.0182, found: 415.0168.

4-(4-Cyanophenyl)-1-[(5-(3,4-dichlorophenyl)furan-2-yl)methylene]thiosemicarbazide (8)

M.p. 202 °C. Yield 82%.

IR v_{max} (cm⁻¹): 3495.01 (N-H stretching), 3147.83, 3010.88 (Aromatic C-H stretching), 2983.88 (Aliphatic C-H stretching), 2227.78 (C \equiv N stretching), 1625.99, 1602.85, 1585.49, 1546.91, 1519.91, 1456.26 (N-H bending, C=N and C=C stretching), 1411.89 (C-H bending), 1330.88, 1273.02, 1219.01, 1176.58, 1138.00, 1082.07, 1029.99 (C-N, C-O stretching and aromatic C-H in plane bending), 981.77, 935.48, 923.90, 867.97, 837.11, 815.89, 792.74, 771.53, 734.88, 696.30 (Aromatic C-H out of plane bending).

 1 H NMR (400 MHz, DMSO- d_6) δ (ppm): 7.24 (d, J = 4.00 Hz, 1H, aromatic proton), 7.32 (d, J = 4.00 Hz, 1H, aromatic proton), 7.69-7.85 (m, 4H, aromatic protons), 8.00-8.08 (m, 3H, aromatic protons), 8.14 (s, 1H, -CH=N), 10.23 (s, 1H, S=C-NH), 12.19 (s, 1H, C=N-NH).

 13 C NMR (100 MHz, DMSO- d_6) δ (ppm): 102.44 (C), 106.71 (CH), 110.37 (CH), 116.24 (CH), 116.93 (C), 118.86 (C), 123.94 (CH), 124.63 (2CH), 125.50 (CH), 131.17 (CH), 129.83 (C), 132.24 (2CH), 132.90 (CH), 143.33 (CH), 149.51 (C), 152.41 (C), 175.08 (C).

HRMS (ESI) (m/z): $[M+H]^+$ calcd. for $C_{19}H_{12}Cl_2N_4OS$: 415.0182, found: 415.0185.

4-(4-Cyanophenyl)-1-[(5-(2,5-dichlorophenyl)furan-2-yl)methylene]thiosemicarbazide (9)

M.p. 201.8 °C. Yield 81%.

IR v_{max} (cm⁻¹): 3290.56 (N-H stretching), 3140.11, 3010.88 (Aromatic C-H stretching), 2968.45 (Aliphatic C-H stretching), 2227.78 (C \equiv N stretching), 1606.70, 1589.34, 1521.84, 1463.97 (N-H bending, C \equiv N and C \equiv C stretching), 1442.75, 1413.82, 1379.10 (C-H bending), 1323.17, 1269.16, 1201.65, 1176.58, 1134.14, 1097.50, 1024.20 (C-N, C-O stretching and aromatic C-H in plane bending), 987.55, 937.40, 914.26, 883.40, 835.18, 804.32, 783.10, 707.88, 675.09 (Aromatic C-H out of plane bending).

Research Article

¹H NMR (400 MHz, DMSO- d_6) δ (ppm): 7.28 (d, J = 4.00 Hz, 1H, aromatic proton), 7.38-7.42 (m, 1H, aromatic proton), 7.58 (d, J = 8.80 Hz, 1H, aromatic proton), 7.71-7.75 (m, 1H, aromatic proton), 7.81 (d, J = 8.80 Hz, 2H, aromatic protons), 8.00-8.04 (m, 1H, aromatic proton), 8.12 (s, 1H, -CH=N), 10.27 (s, 1H, S=C-NH), 12.22 (s, 1H, C=N-NH).

 13 C NMR (100 MHz, DMSO- 4 6) δ (ppm): 102.38 (C), 106.68 (CH), 114.39 (CH), 115.45 (CH), 116.89 (C), 118.84 (C), 124.57 (2CH), 127.24 (CH), 127.77 (CH), 129.00 (CH), 132.30 (2CH, d, 1 5 - 7.7 Hz), 132.60 (CH, d, 1 6 - 22.5 Hz), 133.43 (C), 143.30 (CH), 149.47 (C, d, 1 6 - 3.8 Hz), 155.69 (C), 175.15 (C).

HRMS (ESI) (m/z): $[M+H]^+$ calcd. for $C_{19}H_{12}Cl_2N_4OS$: 415.0182, found: 415.0177.

4-(4-Cyanophenyl)-1-[(5-(4-chloro-2-nitrophenyl)furan-2-yl)methylene]thiosemicarbazide (10)

M.p. 196.8 °C. Yield 85%.

IR v_{max} (cm⁻¹): 3298.28 (N-H stretching), 3124.68 (Aromatic C-H stretching), 2972.31 (Aliphatic C-H stretching), 2222.00 (C \equiv N stretching), 1627.92, 1606.70, 1591.27, 1552.70, 1519.91, 1460.11 (N-H bending, NO₂, C \equiv N and C \equiv C stretching), 1442.75, 1402.25, 1375.25 (C-H bending), 1346.31, 1325.10, 1263.37, 1201.65, 1192.01, 1174.65, 1114.86, 1083.99, 1016.49 (C-N, C-O stretching and aromatic C-H in plane bending), 983.70, 925.83, 914.26, 879.54, 829.39, 783.10, 758.02, 696.30 (Aromatic C-H out of plane bending).

 1 H NMR (400 MHz, DMSO- d_{6}) δ (ppm): 7.13-7.22 (m, 1H, aromatic proton), 7.27 (d, J = 3.20 Hz, 1H, aromatic proton), 7.60-7.65 (m, 1H, aromatic proton), 7.72-7.76 (m, 1H, aromatic proton), 7.83-7.88 (m, 1H, aromatic proton), 7.95-8.08 (m, 3H, aromatic protons), 8.16-8.20 (m, 2H, -CH=N and aromatic proton), 10.08 (s, 1H, S=C-NH), 12.30 (s, 1H, C=N-NH).

 13 C NMR (100 MHz, DMSO- 2 6) δ (ppm): 102.38 (C), 106.67 (CH), 112.77 (CH), 115.41 (CH), 116.89 (C), 118.79 (C), 120.28 (CH), 123.74 (2CH, d, 2 7 = 8.4 Hz), 129.85 (CH), 132.13 (CH, d, 2 7 = 13.7 Hz), 132.45 (2CH), 133.42 (CH), 142.94 (CH), 146.92 (C), 148.33 (C), 150.41 (C), 174.78 (C).

HRMS (ESI) (m/z): $[M+H]^+$ calcd. for $C_{19}H_{12}ClN_5O_3S$: 426.0422, found: 426.0424.

4.2. Biochemistry

4.2.1. Cell culture and drug treatment

Cell lines were obtained from American Type Culture Collection (ATCC). A549 human lung adenocarcinoma and L929 mouse fibroblast cells were cultured and drug treatments were carried out as previously described [22].

4.2.2. MTT assay

MTT assay was performed as previously described in the literature [22] with small modifications [20]. Cisplatin was used as a positive control. The SI values were also calculated according to the formula [23] below:

 $SI = IC_{50}$ for normal cell line / IC_{50} for cancerous cell line

4.2.3. Flow cytometric analyses of apoptosis

After the incubation of A549 cells with compounds 1, 5, 6, 7 and cisplatin at IC₅₀ concentrations, phosphatidylserine externalization, which indicates early apoptosis, was detected using FITC Annexin V Apoptosis Detection Kit (BD Pharmingen, San Jose, CA, USA) on a BD FACSAria flow cytometer for 24 h. Annexin V staining protocol was applied according to the manufacturer's instructions (BD Pharmingen, San Jose, CA, USA) and analyzed by a BD FACSAria flow cytometer using FACSDiva version 6.1.1 software (BD Biosciences, San Jose, CA, USA).

Acknowledgements: This study was supported by Anadolu University Scientific Research Projects Commission under the grant no: 1805S185.

Author contributions: Concept – B.S., A.Ö., M.D.A.; Design – B.S., A.Ö., M.D.A.; Supervision – A.Ö., M.D.A.; Resource – M.D.A.; Materials – B.S., G.A.Ç., M.D.A.; Data Collection and/or Processing – B.S., G.A.Ç., M.D.A.; Analysis and/or Interpretation – G.A.Ç., A.Ö., M.D.A.; Literature Search – B.S., M.D.A.; Writing – B.S., G.A.Ç., A.Ö., M.D.A.; Critical Reviews – B.S., G.A.Ç., A.Ö., M.D.A.

Conflict of interest statement: The authors declared no conflict of interest.

REFERENCES

- [1] Wood SL, Pernemalm M, Crosbie PA, Whetton AD. Molecular histology of lung cancer: from targets to treatments. Cancer Treat Rev. 2015; 41(4): 361-375. [CrossRef]
- [2] Gyoba J, Shan S, Roa W, Bédard ELR. Diagnosing lung cancers through examination of micro-RNA biomarkers in blood, plasma, serum and sputum: a review and summary of current literature. Int J Mol Sci. 2016; 17(4): 494. [CrossRef]
- [3] Li L, Zhu T, Gao Y-F, Zheng W, Wang C-J, Xiao L, Huang MS, Yin JY, Zhou HH, Liu ZQ. Targeting DNA damage response in the radio(chemo)therapy of non-small cell lung cancer. Int J Mol Sci. 2016; 17(6): E839. [CrossRef]
- [4] Ansari J, Shackelford RE, El-Osta H. Epigenetics in non-small cell lung cancer: from basics to therapeutics. Transl Lung Cancer Res. 2016; 5(2): 155-171. [CrossRef]
- Gavrilescu LC, Denkers EY. Apoptosis and the balance of homeostatic and pathologic responses to protozoan infection. Infect Immun. 2003; 71(11): 6109-6115. [CrossRef]
- El-Khattouti A, Selimovic D, Haikel Y, Hassan M. Crosstalk between apoptosis and autophagy: molecular mechanisms and therapeutic strategies in cancer. J Cell Death. 2013; 6: 37-55. [CrossRef]
- Mohammad RM, Muqbil I, Lowe L, Yedjou C, Hsu H-Y, Lin L-T. Broad targeting of resistance to apoptosis in cancer. Semin Cancer Biol. 2015; 35: S78-S103. [CrossRef]
- Kamal A, Faazil S, Malik MS. Apoptosis-inducing agents: a patent review (2010 2013). Expert Opin Ther Pat. 2014; 24(3): 339-354. [CrossRef]
- [9] Wong RSY. Apoptosis in cancer: from pathogenesis to treatment. J Exp Clin Cancer Res. 2011; 30: 87. [CrossRef]
- [10] Pore MM, Hiltermann TJN, Kruyt FAE. Targeting apoptosis pathways in lung cancer. Cancer Lett. 2013; 332(2): 359-368. [CrossRef]
- [11] Moorthy NSHN, Cerqueira NMFSA, Ramos MJ, Fernandes PA. Aryl- and heteroaryl-thiosemicarbazone derivatives and their metal complexes: a pharmacological template. Recent Pat Anti-Cancer Drug Discov. 2013; 8(2): 168-182. [CrossRef]
- [12] Yu Y, Kalinowski DS, Kovacevic Z, Siafakas AR, Jansson PJ, Stefani C, Lovejoy DB, Sharpe PC, Bernhardt PV, Richardson DR. Thiosemicarbazones from the old to new: iron chelators that are more than just ribonucleotide reductase inhibitors. J Med Chem. 2009; 52(17): 5271-5294. [CrossRef]
- [13] Kalinowski DS, Quach P, Richardson DR. Thiosemicarbazones: the new wave in cancer treatment. Future Med Chem. 2009; 1(6): 1143-1151. [CrossRef]
- [14] Mannargudi MB, Deb S. Clinical pharmacology and clinical trials of ribonucleotide reductase inhibitors: is it a viable cancer therapy? J Cancer Res Clin Oncol. 2017; 143(8): 1499-1529. [CrossRef]
- [15] Yee EMH, Brandl MB, Black DS, Vittorio O, Kumar N. Synthesis of isoflavene-thiosemicarbazone hybrids and evaluation of their anti-tumor activity. Bioorg Med Chem Lett. 2017; 27(11): 2454-2458. [CrossRef]
- [16] Wang Y, Gu W, Shan Y, Liu F, Xu X, Yang Y, Zhang Q, Zhang Y, Kuang H, Wang Z, Wang S. Design, synthesis and anticancer activity of novel nopinone-based thiosemicarbazone derivatives. Bioorg Med Chem Lett. 2017; 27: 2360-2363. [CrossRef]
- [17] Pape VFS, Tóth S, Füredi A, Szebényi K, Lovrics A, Szabó P, Wiese M, Szakács G. Design, synthesis and biological evaluation of thiosemicarbazones, hydrazinobenzothiazoles and arylhydrazones as anticancer agents with a potential to overcome multidrug resistance. Eur J Med Chem. 2016; 117: 335-354. [CrossRef]
- [18] Altıntop MD, Atlı Ö, Ilgın S, Demirel R, Özdemir A, Kaplancıklı ZA. Synthesis and biological evaluation of new naphthalene substituted thiosemicarbazone derivatives as potent antifungal and anticancer agents. Eur J Med Chem. 2016; 108: 406-414. [CrossRef]
- [19] Altıntop MD, Sever B, Özdemir A, Kuş G, Oztopcu-Vatan P, Kabadere S, Kaplancikli ZA. Synthesis and evaluation of naphthalene-based thiosemicarbazone derivatives as new anticancer agents against LNCaP prostate cancer cells. J Enzyme Inhib Med Chem. 2016; 31(3): 410-416. [CrossRef]
- [20] Altıntop MD, Temel HE, Sever B, Akalın Çiftçi G, Kaplancıklı ZA. Synthesis and evaluation of new benzodioxolebased thiosemicarbazone derivatives as potential antitumor agents. Molecules 2016; 21(11): E1598. [CrossRef]
- [21] Serra S, Moineaux L, Vancraeynest C, Masereel B, Wouters J, Pochet L, Frédérick R. Thiosemicarbazide, a fragment with promising indolamine-2,3-dioxygenase (IDO) inhibition properties. Eur J Med Chem. 2014; 82: 96-105. [CrossRef]

- [22] Mosmann T. Rapid colorimetric assay for cellular growth and survival: application to proliferation and cytotoxicity assays. J Immunol Methods 1983; 65: 55-63. [CrossRef]
- [23] Zhou H-Y, Dong F-Q, Du X-L, Zhou Z-K, Huo H-R, Wang W-H, Zhan HD, Dai YF, Jing Meng, Sui YP, Li J, Sui F, Zhai YH. Antitumor activities of biscoumarin and dihydropyran derivatives. Bioorg Med Chem Lett. 2016; 26(16): 3876-3880. [CrossRef]

This is an open access article which is publicly available on our journal's website under Institutional Repository at http://dspace.marmara.edu.tr.