In vitro and *in silico* cytotoxicity effects of *Zanthoxylum simulans* Hance. fruit bark extract against gastric cancer cell lines

Bui Thanh TUNG ¹* (D), Nguyen Thi THUY¹ (D), Le Thi HUONG¹ (D), Tran Hoang MAI¹ (D), Vu Manh HA² (D), Do Thi Hong KHANH (D)

- ¹ Department of Pharmacology, Faculty of Pharmacy, VNU University of Medicine and Pharmacy, Vietnam National University Hanoi, Ha Noi, Vietnam
- ² Department of Pharmacology, Faculty of Pharmacy, Phenikaa University, Ha Noi, Vietnam
- *Corresponding Author. E-mail: <u>tungbt.ump@vnu.edu.vn</u> Tel. +84-904429676.

Received: 05 April 2023 / Revised: 20 June 2023 / Accepted: 21 June 2023

ABSTRACT: *Zanthoxylum simulans* Hance. is known in folklore as a spicy herb, commonly used to treat cold-induced diseases in the body. In this study, we evaluated the cytotoxic effects of *Zanthoxylum simulans* fruit bark extract. Samples of *Zanthoxylum simulans* were extracted with 70% ethanol and subsequently fractionated with n-Hexane, ethyl acetate (EtOAc), and n-butanol (n-BuOH) solvents. To evaluate the in vitro cytotoxic effect, we performed SRB (Sulforhodamine ^B) assay on the cell lines of human gastric MKN-7. In this study, we also used molecular docking method to evaluate the inhibition abilities of MAPK1 and AKT1 receptors of 120 compounds in *Zanthoxylum simulans* Hance. The *in vitro* cytotoxic results showed that the n-Hexane total extract had the strongest cytotoxicity effects on gastric cancer cells with an IC₅₀ of 23.65±1.75 mg/ml. The results cytotoxic effects on MNK-7 gastric cancer cells of these fractions showed that the EtOAc and EtOH fractions exhibited activity with IC₅₀ values of 35.61±2.90 and 50.67±3.82 µg/mL, respectively; while the BuOH and H₂O fractions showed no activity. The molecular docking results showed five compounds that inhibit both MAPK1 and AKT1 targets including Simulanoquinoline, N-acetylanonaine, N-acetyldehydroanonaine, Oxyavicine, and Benzosimuline. Therefore, our results showed that *Zanthoxylum simulans* fruit bark extract has a strong cytotoxicity effect on gastric cancer cells. *In vivo* studies of these potential compounds should be carried out to become anti-cancer drugs in the future.

KEYWORDS: Zanthoxylum simulans Hance.; cytotoxicity; molecular docking; gastric cancer cells; MAPK1; AKT1

1. INTRODUCTION

Cancer is a complex disease caused by genetic alterations such as gene mutations or changes in the expression of cancer-related genes or tumor suppressor genes associated with chromosomes [1]. Gastric cancer, an aggressive form of digestive system tumours, has the third highest lethality and fourth highest morbidity in all cancers worldwide. It is a complex disease with multiple genetic and environmental factors involved in its development and progression.

MAPK1 is a member of the mitogen-activated protein kinase (MAPK) family, which is involved in various cellular processes, including cell proliferation, differentiation, survival, and apoptosis [2]. It is activated by a cascade of phosphorylation events initiated by upstream signaling molecules, such as growth factors, cytokines, and stress stimuli. Several studies have shown that MAPK1 is upregulated in gastric cancer tissues compared to normal gastric tissues. In addition, its expression levels correlate with the stage and grade of the disease, suggesting that it may play a role in gastric cancer progression. One of the mechanisms by which MAPK1 promotes gastric cancer is through the regulation of cell proliferation and apoptosis [3].

MAPK1 activation stimulates the expression of genes involved in cell cycle progression, such as cyclin D1 and c-Myc, while inhibiting the expression of pro-apoptotic genes, such as Bax and caspase-3. This leads

Tung BT, Thuy NT, Huong LT, Mai TH, Ha VM, Khanh DTH. In vitro and in silico cytotoxicity effects of Zanthoxylum simulans Hance. fruit bark extract against gastric cancer cell lines. J Res Pharm. 2024; 28(1): 110-125.

to increased cell proliferation and decreased apoptosis, which are hallmarks of cancer cells. Targeting MAPK1 signaling pathway has emerged as a promising strategy for the treatment of gastric cancer [4].

AKT1, also known as protein kinase B (PKB), is a serine/threonine protein kinase that plays a crucial role in regulating various cellular processes, including cell proliferation, differentiation, survival, and metabolism [5]. Dysregulation of AKT1 signaling has been implicated in various types of cancer, including gastric cancer. Several studies have reported that AKT1 expression is upregulated in gastric cancer tissues compared to normal gastric tissues. In addition, high AKT1 expression levels have been associated with a poor prognosis in gastric cancer patients, suggesting that AKT1 may be involved in gastric cancer progression. One of the mechanisms by which AKT1 promotes gastric cancer is through its effects on cell proliferation and apoptosis. AKT1 activation leads to the upregulation of cell cycle regulators, such as cyclin D1 and c-Myc, and the downregulation of pro-apoptotic genes, such as Bax and caspase-3. This promotes cell proliferation and inhibits apoptosis, which are hallmarks of cancer cells [6]. AKT1 also plays a role in regulating the invasive and metastatic potential of gastric cancer cells. AKT1 activation stimulates the expression of matrix metalloproteinases (MMPs), which are enzymes that degrade the extracellular matrix and facilitate cancer cell invasion and migration. Targeting AKT1 signaling pathway has emerged as a promising therapeutic approach for the treatment of gastric cancer [7].

Molecular docking is a modeling technique to predict the favorable position and configuration that the substrate molecule can bind to a protein. It can predict the interaction of the compound with acid amine in the protein target. The substrate molecule is displaced through space and encircles the protein molecule to find the position with the most negative binding energy. In most cases of cancer, chemotherapy or radiation therapy is the primary treatment option [8]. However, radiation therapy often causes many side effects, chemotherapy can be costly, and the five-year survival rate for patients remains limited. One approach that is currently receiving special attention is the use of whole-plant extracts or compounds extracted from medicinal plants to treat cancer [9]. Medicinal plants are a readily available source of materials, cost-effective, have good efficacy, and have few undesirable side effects. Zanthoxylum simulans Hance. is a wild-growing plant that is found in tropical climates. It is commonly found in eastern China, Taiwan, North Korea, Cambodia, and Laos. In Vietnam, this plant is found in the central region. Chemical analysis of the Xuyen tieu plant has shown that it contains main compounds belonging to the groups of alkaloids, amides, lignans and neolignans, coumarins, peptides, terpenoids, and flavonoids [10-12]. Pharmacological studies have shown that Zanthoxylum simulans has many important effects, including anti-inflammatory, antibacterial, antiparasitic, antiviral, antioxidant, anticancer/antitumor, and cytotoxic effects [10]. In this study, we conducted to evaluate the in vitro cytotoxic effects of Zanthoxylum simulans fruit bark extract against gastric cancer cell lines. We also performed in silico molecular docking, and absorption, distribution, metabolism, excretion, and toxicity (ADMET) studies to demonstrate the probable interactions between promised compounds in Zanthoxylum simulans with MAPK1 and AKT1 and their ADMET profile.

2. RESULTS

2.1. In vitro cytotoxicity effects on gastric cancer cells of Zanthoxylum simulans

In vitro cytotoxicity effects on gastric cancer cells of the total ethanolic extract and fractions of *Zanthoxylum simulans* fruit bark were demonstrated in Table 1.

Table 1. In vitro cytotoxicity effects of the total ethanolic extract and fractions of Zanthoxylum simulans fruit bark on gastric cancer cells

| MNK-7 | EtOH | n-Hexan | EtOAc | BuOH | H ₂ O | Ellipticine |
|------------------|------------|------------|------------|------|------------------|-------------|
| IC ₅₀ | 50.67±3.82 | 23.65±1.75 | 35.61±2.90 | >100 | >100 | 0.40±0.03 |

From Table 1, the positive control drug Ellipticine showed significant toxicity against the human gastric cancer cell line with an IC_{50} of 0.40±0.03. The n-Hexane total extract displayed cytotoxicity against the human

gastric cancer cell line with an IC₅₀ of 23.65±1.75 mg/ml. Results in Table 1 also showed that the EtOAc and EtOH fractions demonstrated activity with IC₅₀ values of 35.61±2.90 and 50.67±3.82 μ g/mL, respectively. The BuOH and H₂O fractions did not exhibit activity against the tested cell line.

2.2. Evaluation of the docking model

Before screening compounds, the co-crystallized ligands (Ipatasertib and Ravoxertinib) were redocked to the active site of the target by Chimera. We received the root mean square deviation (RMSD) of AKT1 and MAPK1 are 0.432 Å and 0.464 Å, respectively (Figure 1 & Figure 2). All RMSD values are less than 1.5 Å proving that molecular docking results to the target are reliable.

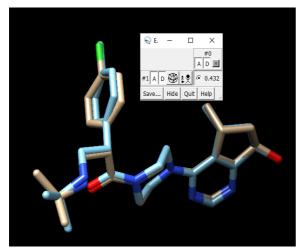


Figure 1. Co-crystallized ligand redock results of AKT1

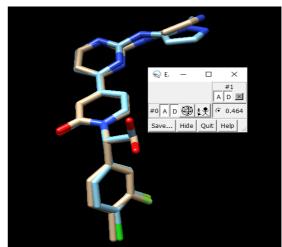


Figure 2. Co-crystallized ligand redock results of MAPK1

2.3. Molecular docking of compounds to the target protein

After preparing the proteins, we docked 120 natural compounds from *Zanthoxylum simulans* Hance. to screen inhibitory activity in AKT1 and MAPK1 targets. The result is shown in Table 2.

| No. | Name | PubChemID | Binding energy (kcal/mol) | | |
|-----|-------------------------|-----------|---------------------------|-------|--|
| | | | AKT1 | MAPK1 | |
| 1 | Zanthosimuline | 5315426 | -8.5 | -8.2 | |
| 2 | Huajiaosimuline | 5318093 | -8.9 | -8.2 | |
| 3 | Simulanoquinoline | 5321314 | -9.5 | -10.3 | |
| 4 | N-acetylanonaine | 6453733 | -9.6 | -9.6 | |
| 5 | Chelerythrine | 2703 | -8.6 | -8.7 | |
| 6 | Norchelerythrine | 443719 | -8.4 | -8.6 | |
| 7 | Bocconoline | 181121 | -8.3 | -8.6 | |
| 8 | Skimmianine | 6760 | -7.4 | -6.7 | |
| 9 | N-acetylnornuciferine | 101630664 | -8.1 | -8.8 | |
| 10 | Arnottianamide | 3085181 | -8.5 | -8.1 | |
| 11 | Decarine | 179640 | -9.0 | -8.6 | |
| 12 | N-acetyldehydroanonaine | 5315739 | -9.8 | -9.1 | |
| 13 | Simulansine | 5321315 | -8.7 | -8.1 | |
| 14 | Oxychelerythrine | 147279 | -9.0 | -8.8 | |
| 15 | Dihydrochelerythrine | 485077 | -8.9 | -8.5 | |
| 16 | Zanthobisquinolone | 54688597 | -9.0 | -8.4 | |

Table 2. The docking results of 120 compounds and reference compounds with AKT1 and MAPK1.

Tung et al. Cytotoxicity effects of Z. simulans Hance. fruit bark extract on gastric cancer

Research Article

| 17 | γ-fagarine | 107936 | -7.2 | -6.6 |
|----------|--------------------------------|------------------------|--------------|--------------|
| 18 10 | Toddaquinoline Dictamnine | 11390791 | -8.6 | -8.4 -6.6 |
| 19 20 | 4-methoxy-2-quinolone | 68085 600167 | -7.1 -6.4 | -6.3 |
| 20 21 | Benzophenanthridine | 12407248 | -8.7 | -8.7 |
| 21 | Noravicine | 14037817 | -0.7 -9.1 | -9.6 |
| | Rhoifoline B | | -9.1 -9.5 | |
| 23 24 | 8-methoxyisodecarine | 377308819 136854509 | -9.3 -9.1 | -9.3 -8.7 |
| 24 25 | Dihydronitidine | 99641 | -9.1 -9.4 | -8.5 |
| 25 26 | 5,6-dihydro-6-methoxynitidine | 38845 | -7.8 | -8.8 |
| 20 27 | 6-acetonyldihydronitidine | 101664489 | -7.8 -9.0 | -8.8 -7.9 |
| 27 | 6-acetonyldihydroavicine | 101004489 | -9.0 | -9.5 |
| 20 29 | 6-acetonyldihydrochelerythrine | 185516 | -8.9 | -7.9 |
| 30 | 8-hydroxydihydrochelerythrine | 15940321 | -8.9 | -8.8 |
| 31 | Ethoxychelerythrine | 160921 | -7.8 | -7.6 |
| 32 | Oxynitidine | 97597 | -9.5 | -9.1 |
| 33 | Oxyavicine | 12313849 | -9.8 | -9.5 |
| 34 | Isoarnottianamide | 14189418 | -9.8 | -7.2 |
| 35 | Integriamide | 155897 | -8.9 | -8.2 |
| 36 | Citronellol | 8842 | -4.7 | -4.9 |
| 37 | Spathulenol | 92231 | -7.5 | -7.0 |
| 38 | Robustine | 164950 | -7.1 | -6.9 |
| 39 | N-methylflindersine | 72819 | -8.0 | -7.9 |
| 40 | Liriodenine | 10144 | -9.2 | -9.2 |
| 41 | Hinokinin | 442879 | -9.0 | -8.6 |
| 42 | Lysicamine | 122691 | -8.7 | -8.4 |
| 43 | Simulenoline | 5321316 | -8.7 | -8.3 |
| 44 | Peroxysimulenoline | 101936038 | -9.1 | -8.2 |
| 45 | Benzosimuline | 5321951 | -9.4 | -9.3 |
| 46 | Edulitine | 826073 | -6.6 | -6.2 |
| 47 | Arborinine | 5281832 | -8.2 | -7.6 |
| 48 | Scoparone | 8417 | -6.6 | -6.4 |
| 49 | β-amyrone | 12306160 | -8.3 | -8.0 |
| 50 | β-amyrin | 73145 | -8.5 | -8.1 |
| 51 | β-sitosterol | 222284 | -7.3 | -7.0 |
| 52 | β-sitostenone | 5484202 | -7.9 | -7.3 |
| 53 | Tetracosyl ferulate | 14238617 | -6.7 | -6.6 |
| 54 | Zanthobungeanine | 5315422 | -8.0 | -7.6 |
| 55 | Pyrrolezanthine | 636825 | -6.7 | -5.8 |
| 56 | (-)-Simulanol | 636826 | -7.9 | -7.7 |
| 57 | Zanthopyranone | 10419590 | -5.2 | -4.7 |
| 58 | Sinapic aldehyde | 5280802 | -5.7 | -5.5 |
| 59 | Vanillic acid | 8468 | -6.0 | -5.4 |
| 60 | Syringic acid | 10742 | -5.8 | -5.5 |
| 61 | Isofraxidin | 5318565 | -6.7 | -6.4 |
| 62 | (-)-Balanophonin | 23252258 | -7.9 | -8.2 |
| 63 | haplopine | 5281846 | -7.5 | -6.7 |
| 64 | (+)-Platydesmine | 6451457 | -7.9 | -7.4 |

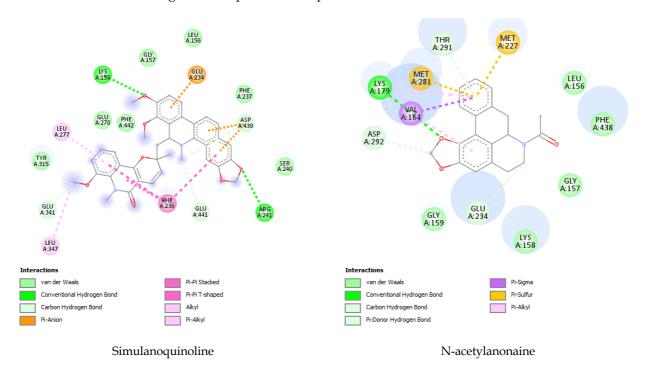
| arch | Article |
|------|---------|
| arcn | Article |

| 65 | (-)-Syringaresinol | 11604108 | -7.9 | -7.9 |
|-----|-------------------------------|-----------|------|------|
| 66 | Flindersine | 68230 | -8.0 | -8.0 |
| 67 | Zanthodioline | 78384601 | -8.0 | -7.7 |
| 68 | Scopoletin | 5280460 | -6.5 | -6.4 |
| 69 | (+)-abscisic acid | 5280896 | -7.7 | -6.9 |
| 70 | Caryophyllene oxide | 1742210 | -6.5 | -6.3 |
| 71 | Isobutyl acetate | 8038 | -4.6 | -4.0 |
| 72 | Kobusin | 182278 | -8.6 | -8.2 |
| 73 | (+)-Fargesin | 5320622 | -8.7 | -8.3 |
| 74 | Epieudesmin | 7000209 | -8.3 | -8.0 |
| 75 | Simulansamide | 5321312 | -7.8 | -7.3 |
| 76 | a-pinene | 6654 | -5.2 | -5.3 |
| 77 | Camphene | 6616 | -5.0 | -5.4 |
| 78 | β-pinene | 14896 | -5.3 | -5.1 |
| 79 | Sabinene | 18818 | -4.8 | -5.2 |
| 80 | Isoamyl acetate | 31276 | -4.5 | -4.1 |
| 81 | β-myrcene | 31253 | -4.9 | -4.8 |
| 82 | a-terpinene | 7462 | -5.9 | -5.7 |
| 83 | Limonene | 22311 | -5.7 | -5.6 |
| 84 | β-phellandrene | 11142 | -5.8 | -5.5 |
| 85 | 1,8-cineole | 2758 | -4.8 | -5.5 |
| 86 | (Z)-β-ocimene | 5320250 | -5.0 | -4.9 |
| 87 | γ-terpinene | 348276756 | -5.9 | -5.6 |
| 88 | (E)-β-ocimene | 5281553 | -5.3 | -5.2 |
| 89 | p-cymene | 7463 | -5.9 | -5.7 |
| 90 | Terpinolene | 11463 | -6.0 | -5.8 |
| 91 | (Z)-3-tridecen-1-yne | 5367347 | -5.4 | -4.5 |
| 92 | Heptyl acetate | 8159 | -4.6 | -4.5 |
| 93 | 3,4-dimethyl-2,4,6-octatriene | 5371124 | -5.5 | -5.3 |
| 94 | (E)-sabinene hydrate | 6430763 | -5.3 | -5.7 |
| 95 | Linalyl acetate | 8294 | -5.2 | -5.1 |
| 96 | N-octanol | 957 | -4.1 | -4.0 |
| 97 | Linalyl formate | 61040 | -5.1 | -4.6 |
| 98 | 4-terpineol | 11230 | -5.8 | -5.2 |
| 99 | β-caryophyllene | 5281515 | -6.9 | -6.5 |
| 100 | Sabina ketone | 92784 | -5.1 | -4.9 |
| 101 | a-humulene | 5281520 | -6.7 | -6.2 |
| 102 | a-terpineo | 17100 | -6.3 | -5.9 |
| 103 | 3-thujen-2-ol | 561871 | -5.5 | -5.5 |
| 104 | α-terpinyl acetate | 111037 | -6.3 | -5.9 |
| 105 | Germacrene D | 5317570 | -6.9 | -6.9 |
| 106 | Neryl acetate | 1549025 | -5.6 | -5.1 |
| 107 | Zingiberene | 92776 | -6.2 | -6.7 |
| 108 | Geranyl acetate | 1549026 | -5.6 | -5.5 |
| 109 | δ-cadinene | 8145817 | -7.5 | -7.0 |
| 110 | Allethrolone | 11083 | -5.9 | -5.4 |
| 111 | Geraniol | 637566 | -4.9 | -4.7 |
| 112 | Elemol | 92138 | -6.1 | -5.8 |

Tung et al. Cytotoxicity effects of Z. simulans Hance. fruit bark extract on gastric cancer Journal of Research in Pharmacy Research Article

| 113 | Allo-ocimene | 5368821 | -5.4 | -5.2 |
|-----|------------------|---------|------|------|
| 114 | a-thujene | 17868 | -5.5 | -5.2 |
| 115 | a-fenchol | 439711 | -5.5 | -5.7 |
| 116 | Trans-2-Pinanol | 1268143 | -5.2 | -5.5 |
| 117 | Camphor | 2537 | -5.0 | -5.2 |
| 118 | Camphene hydrate | 101680 | -4.8 | -5.5 |
| 119 | Isoborneol | 6321405 | -5.0 | -5.0 |
| 120 | Carvone | 7439 | -6.0 | -5.7 |
| 121 | Ipatasertib | | -8.6 | |
| 122 | Ravoxertinib | | | -8.0 |
| | | | | |

Therefore, in this study, we compared the binding energies of potential compounds with positive controls (Ipatasertib and Ravoxertinib) to evaluate the inhibition of AKT1 and MAPK1 ability. Based on Table 2, there are 29 natural compounds with the most negative binding energy with AKT1 than positive control Ipatasertib, 37 natural compounds with the most negative binding energy with MAPK1 than positive control Ravoxertinib. From these results, the five strongest natural compounds that inhibit all both targets AKT1 and MAPK1 receptors were Simulanoquinoline (3), N- acetylanonaine (4), N-acetyldehydroanonaine (12), Oxyavicine (33), Benzosimuline (45). The ligand-amino acid interactions between them and the AKT1 and MAPK1 receptors (Figure 3 & Figure 4) show mainly interactions with II-bonding and hydrogen bonding. The amino acids of these both targets with 5 potential compounds are shown in Table 3.



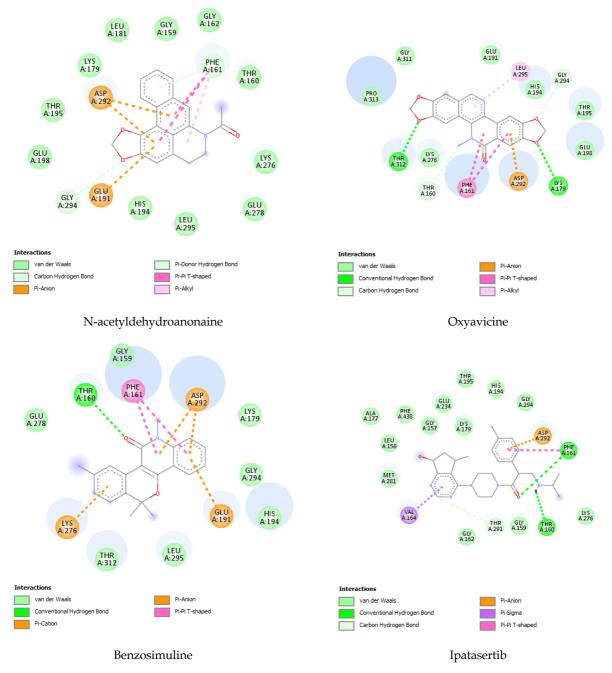
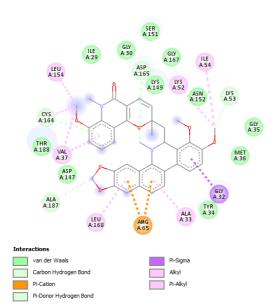
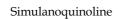
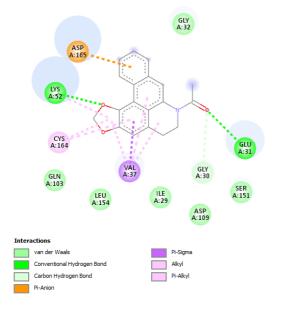
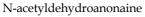


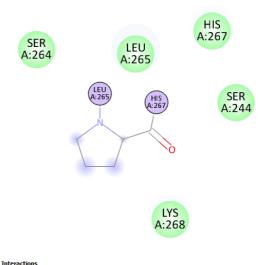
Figure 3. Interactions between 5 compounds and Ipatasertib with AKT1







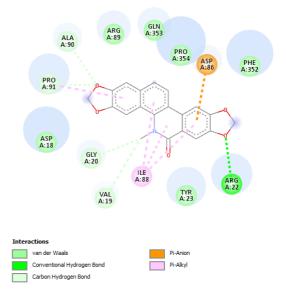




N-acetylanonaine

Covalent bond

van der Waals



Oxyavicine

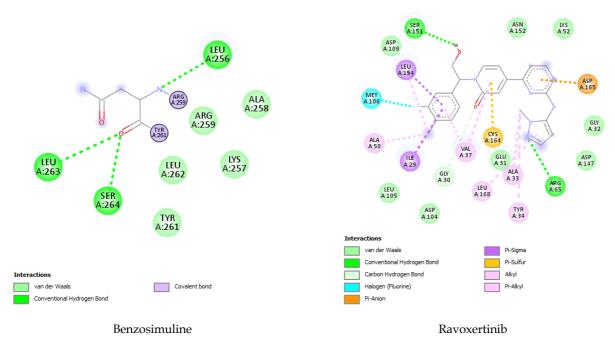


Figure 4. Interactions between 5 compounds and Ravoxertinib with MAPK1

| Compounds | The important amino acids binding | | | | |
|-------------------------------|-------------------------------------|-------------------------------------|--|--|--|
| compounds | AKT1 | MAPK1 | | | |
| | LYS 158, LEU 227, GLU 234, PHE 236, | GLY 32, ALA 33, VAL 37, LYS 52, | | | |
| Simulanoquinoline | ARG 241, GLU 341, LEU 347, ASP 439, | LYS 53, ILE 54, ARG 65, LEU 154, | | | |
| | GLU 441 | CYS 164, ASP 165, LEU 168, ALA 187 | | | |
| N | VAL 164, LYS 179, MET 227, GLU 234, | LEU 265, HIS 267 | | | |
| N-acetylanonaine | MET 281, THR 291, ASP 292 | | | | |
| NI a aatal dahar dugan anaina | | GLY 30, GLU 31, VAL 37, LYS 52, | | | |
| N-acetyldehydroanonaine | PHE 161, GLU 191, ASP 292, GLY 294 | CYS 164, ASP 165 | | | |
| Organisina | THR 160, PHE 161, LYS 179, ASP 292, | VAL 19, GLY 20, ARG 22, ASP 86, ILE | | | |
| Oxyavicine | GLY 294, LEU 295, THR 312 | 88, ALA 90, PRO 91 | | | |
| Description 11 and | THR 160, PHE 161, GLU 191, LYS 276, | LEU 256, ARG 259, TYR 261, LEU 263, | | | |
| Benzosimuline | ASP 292 | SER 264 | | | |
| The stars and 'L | THR 160, PHE 161, GLY 162, VAL 164, | | | | |
| Ipatasertib | ASP 292 | | | | |
| | | ILE 29, GLY 30, ALA 33, TYR 34, VAL | | | |
| Descent of the th | | 37, ALA 50, ARG 65, MET 106, SER | | | |
| Ravoxertinib | | 151, LEU 154, CYS 164, ASP 165, LEU | | | |
| | | 168 | | | |

| Table 3. Amino acids of the 5 | potential compounds that | bind to AKT1 and MAPK1 receptors. |
|-------------------------------|--------------------------|-----------------------------------|
| | | |

Comparing the interactions of these 5 compounds and AKT1-targeted positive controls, it can be seen that the ligand amino acid binding of most compounds are similar. Those are THR 160, PHE 161, ASP 292. For the MAPK1 target, these compounds also have relatively low binding energies to this enzyme through similar amino acids: ASP 165, GLY 30, ALA 33 (Table 3).

2.4. Lipinski's rule of five

Compounds are considered to be "drug-like" when they have at least 2 of the 5 criteria of Lipinski's 5criteria rule: Molecular mass (MW) below 500 Daltons; high lipophilicity (expressed as LogP less than 5); less than 5 hydrogen bond donors (HBD); less than 10 hydrogen bond acceptors (HBA1) and molar refractivity (MR) should be between 40-130. Table 4 showed that all 5 compounds satisfy Lipinski's 5-criteria rule. Next, these compounds were further evaluated for their pharmacokinetic-toxicological properties by predicting ADMET.

| Name | MW | HBD | HBA1 | LogP | MR | Drug- likeness |
|-------------------------|-------|-----|------|--------|--------|-------------------|
| Simulanoquinoline | 618.0 | 0 | 9 | 6.8751 | 176.33 | Yes |
| N-acetylanonaine | 307.0 | 0 | 4 | 3.0840 | 85.5 | Yes |
| N-acetyldehydroanonaine | 305.0 | 0 | 4 | 3.6307 | 89.3 | Yes |
| Oxyavicine | 347.0 | 0 | 6 | 3.5541 | 94.36 | Yes |
| Benzosimuline | 305.0 | 0 | 3 | 4.1050 | 91.95 | Yes |

Table 4. The result of Lipinski's rule five.

2.5. Prediction of ADMET profile

To evaluate the effectiveness of the above 5 substances, we continuously evaluate the pharmacokinetic and toxicological parameters (ADMET) through pkCSM. Table 5 is the ADMET prediction result.

In the absorption process, human colon adenocarcinoma-2 cell line (Caco2) permeability and human intestinal absorption (HIA) are crucial benchmarks to determine the entire bioavailability of a drug [13, 14]. Caco2 membrane permeability (log Papp in 10^{-6} cm/s) with a higher value than 0.9 is said to have good permeability. Table 5 indicates that five compounds have good permeability to the CaCo2 cell membrane, with value log Papp in 10⁻⁶ cm/s in range from 1.022 to 1.937. Five compounds also showed good absorption through the human intestine with absorption percentages from 97.303 to reach 100%. For distribution, a logBBB value greater than 0.3 is said to be well absorbed across the blood-brain barrier and less than -1 is considered not to cross the blood-brain barrier. The results showed that five compounds had little permeability through the blood-brain barrier except Benzosimuline has logBBB is 0.285. In terms of metabolism, the cytochrome P450 system is an important enzyme system in drug metabolism in the liver, with two important CYPs, CYP3A4 and CYP2D6. Here, all of the above compounds are substrates of CYP3A4, indicating that they can be metabolized in the liver. Furthermore, Benzosimuline and Oxyavicine may inhibit CYP3A4. In terms of elimination and toxicity, all compounds are capable of renal elimination. The ADMET prediction results also show that simulanoquinoline gives the best results with predictive properties of no AMES toxicity and no liver toxicity and could not cause skin irritation. Four compounds including N-acetyldehydroanonaine, Nacetylanonaine, Benzosimuline and Oxyavicine may have AMES toxicity and Hepatotoxicity but not cause skin sensitisation.

| Table 5. Pharmacokine | etic and toxicol | ogical predictior | ı results. |
|-----------------------|------------------|-------------------|------------|
| | | | |

| Properties | Simulanoquinoline | N-acetyldehydroanonaine | N-acetylanonaine | Oxyavicine | Benzosimuline |
|--|-------------------|-------------------------|------------------|------------|---------------|
| | | Absorption | | | |
| Water solubility (log mol/l) | -4.743 | -4.831 | -3.764 | -5.507 | -4.647 |
| Caco2 permeability (log Papp in 10 ⁻⁶ cm/s) | 1.022 | 1.719 | 1.937 | 1.284 | 1.532 |
| Intestinal absorption (human) (%) | 100 | 99.256 | 97.303 | 100 | 99.096 |
| |] | Distribution | | | |
| VDss (human) (log L/kg) | -0.899 | 0.353 | 0.436 | -0.165 | 0.469 |
| BBB permeability (log BB) | -1.238 | -0.15 | -0.149 | -0.389 | 0.285 |
| | | Metabolism | | | |
| CYP2D6 substrate | No | No | No | No | No |
| CYP3A4 substrate | Yes | Yes | Yes | Yes | Yes |
| CYP2D6 inhibitor | No | No | No | No | No |
| CYP3A4 inhibitor | No | No | No | Yes | Yes |
| | | Excretion | | | |
| Total clearance (log ml/min/kg) | 0.373 | 0.14 | 0.136 | 0.08 | 0.307 |
| | | Toxicity | | | |
| AMES toxicity | No | Yes | Yes | Yes | Yes |
| Hepatotoxicity | No | Yes | Yes | Yes | Yes |
| Skin sensitisation | No | No | No | No | No |

3. DISCUSSION

We conducted the SRB method to evaluate the cytotoxicity of different fractions of Zanthoxylum simulans fruit bark extract on gastric cancer cells. The total n-hexane extract exhibited inhibitory activity on MNK-7 gastric cancer cells line with an IC₅₀ value of $23.65\pm1.75 \ \mu g/mL$. The EtOAc and EtOH fractions showed cytotoxicity activities with IC₅₀ values of and 35.61 ± 2.90 and $50.67\pm3.82 \,\mu$ g/mL, respectively. Our results are consistent with previous studies. Chao Wang and colleagues extracted acridone alkaloids from Zanthoxylum simulans fruit root bark and evaluated their inhibitory effects on prostate cancer PC-3M and LNCaP cells as well as *Plasmodium falciparum* parasites. These authors showed that five acridone alkaloids, including normelicopidine, normelicopine, melicopine, melicopidine, and melicopicine, exhibited inhibitory effects on cancer cells and anti-malarial activity in vitro [15]. In another study, Nguyen Bich Thu and colleagues screened medicinal plants for their anti-cancer effects and found that methanol extracts leaves and seeds of Zanthoxylum simulans inhibited HepG2 cancer cells with IC₅₀ values of 7.1 \pm 0.1 and 28.5 \pm 4.3 μ g/mL, respectively. Furthermore, they showed that the extracts also showed inhibitory effects on other cancer cell lines, such as lung cancer A549, liver cancer Huh-7, fibrosarcoma HT1080, and breast cancer MCF-7 [16]. Recently, Yong-Qiang Tian showed that chelerythrine, a compound in Zanthoxylum simulans exhibited stronger inhibitory effects on gastric cancer cells than Cerdulatinib, a positive control. Chelerythrine inhibited cell adhesion, migration, invasion, and induced programmed cell death in AGS gastric cancer cells, reduced the expression of estrogen receptors (ER- α 36, ER- α 66, and ER- β 1) and Src proto-oncogene [17].

In this study, we evaluated the inhibitory ability of 120 compounds in *Zanthoxylum simulans* Hance. with both MAPK1 and AKT1 targets. After performing molecular docking, we obtained five compounds showing high inhibitory ability on these 2 targets, with lower binding energy than positive controls and druglike properties, including Simulanoquinoline, N-acetyldehydroanonaine, N-acetylanonaine, Benzosimuline, and Oxyavicine. Simulanoquinoline is a benzophenanthridine alkaloid. The simulanoquinoline dimeric profile was first identified in Zanthoxylum simulans in 1993 by Shwu-Jen and Ih-Sheng [18]. This compound is found in many plants of the genus Zanthoxylum such as Zanthoxylum simulans Hance., and Zanthoxylum rhetsa (Roxb.) DC [19]. Similar to simulanoquinoline, N-acetyldehydroanonaine is also found in many plants such as Zanthoxylum nitidum, Zanthoxylum simulans [11, 20]. The genus Zanthoxylum has been studied with many biological effects such as anti-cancer cell proliferation, cytotoxicity, anti-inflammatory, antioxidant, antibiotic, hepatoprotective, and antiviral,... mainly due to the presence of alkaloids and essential oils [21]. In our study, Simulanquinoline showed the ability to inhibit both AKT1 and MAPK1 targets simultaneously with docking scores of -9.5 and -10.3 kcal/mol, respectively. Meanwhile, N-acetyldehydroanonaine has docking scores of -9.6 with both targets. Besides, these two compounds also show the potential to become an anti-cancer drug when satisfying the criteria of Lipinski's rule of five, predicting a positive AMET (well-absorbed, less metabolized by the liver, eliminated by the kidney). Therefore, further studies need to be conducted to more accurately assess the drug potential of this compound.

N-acetylanonaine is an alkaloid, found in many plants such as *Zanthoxylum simulans*, and *Magnolia kachirachirai* [22, 23]. This compound has been reported for its complete inhibitory ability on platelet aggregation induced by arachidonic acid and collagen as well as antimicrobial activity in both bacteria and fungi such as *C. albicans* and *S. aureus* [23, 24]. In our study, N-acetylanonaine exhibited the ability to inhibit both AKT1 and MAPK1 targets with docking scores of -9.8 and -9.1 kcal/mol, respectively. Besides, this compound also shows the potential to become an anti-cancer drug when satisfying the criteria of Lipinski's rule of five, predicting a positive AMET (non-skin sensitisation, well-absorbed, less metabolized by the liver, eliminated by the kidney). Although N-acetylanonaine may have AMES toxicity and hepatotoxicity, further studies need to be evaluated its potential to become an anti-gastric cancer drug in the future.

Oxyavicine is an alkaloid with antinociceptive and anti-inflammatory abilities [25]. In this study, oxyavivine inhibits both AKT1 and MAPK1 targets with docking scores of -9.8 and -9.1 kcal/mol, respectively through binding acid amines similar to positive control. Moreover, this compound also showed the potential to become a drug through drug-likeness, and positive pharmacology. Therefore, oxyavicine is a potential compound to treat gastric cancer in the future.

Benzosimuline is anti-platelet aggregation activity that has been isolated from the plant *Zanthoxylum simulans* [26]. Our results showed that benzosimuline strongly inhibits both AKT1 and MAPK1 with a free binding energy of -9.4 and -9.3 kcal/mol, which is much more negative than the positive controls, Ipatasertib and Ravoxertinib. The ADMET profile suggested that this compound has renal elimination and is well-absorbed. Therefore, we can see the great potential of benzosimuline in inhibiting gastric cancer cells.

4. CONCLUSION

Our study evaluated the *in vitro* anticancer effects of *Zanthoxylum simulans* fruit bark extract on gastric cancer cells line. The total n-Hexan extract and two fractions, EtOAc and EtOH, showed the strongest activity against gastric cancer cells line with IC₅₀ values of 23.65 ± 1.75 ; 35.61 ± 2.90 and $50.67\pm3.82 \mu g/mL$, respectively. Through the results of screening 120 compounds from *Zanthoxylum simulans*, we have found the five most potential compounds, including Simulanoquinoline, N-acetyldehydroanonaine, N-acetylanonaine, Benzosimuline and Oxyavicine. These compounds could simultaneously inhibit MAPK1 and AKT1 targets with the most negative binding energy, drug-likeness and have the best ADMET results including good absorption, renal elimination, and low toxicity. Therefore, *in vitro* and *in vivo* studies are needed to develop these potential compounds as anti-cancer drugs.

5. MATERIALS AND METHODS

5.1. Plant material

The fruit bark of *Zanthoxylum simulans* was purchased in December 2022 from Hanoi Vietnam. The plant samples were authenticated, and a voucher specimen (No: DLPC23UMP-VNU) has been deposited at the Department of Pharmacology, University of Medicine and Pharmacy, Vietnam National University, Hanoi, Vietnam.

5.2. Extraction

The dried fruit bark of *Zanthoxylum simulans* (500 g) was extracted with 70% EtOH at room temperature, 3 times x 3 days with a ratio of medicinal plants/solvent 1:10 (kg/l). The combined extracts were filtered and evaporated under reduced pressure to yield a green residue (23.7 g), which was suspended in water and successively partitioned with organic solvents, then concentrated to yield three extracts of *n*-hexane (1.2 g), ethyl acetate (EtOAc, 7.6 g), *n*-butanol (4.7 g), and a water-soluble layer (8.1 g).

5.3. Cytotoxicity assay

The SRB assay is used for cell density determination, based on the measurement of cellular protein content [27]. The method described here has been optimized for the toxicity screening of compounds to adherent cells in a 96-well format. After an incubation period, cell monolayers were fixed with 10% (wt/vol) trichloroacetic acid and stained for 30 min; then, excess dye was removed by washing the cells repeatedly with 1% (vol/vol) acetic acid. The protein-bound dye was dissolved in 10 mM Tris base solution for optical density (OD) determination at 540 nm using a microplate reader. Ellipticine was used as control positive. The cytotoxicity activity was expressed as IC_{50} values.

5.4. Molecular docking study

Compounds were docked into protein binding cavities using Autodock Tools software.

5.4.1. Preparation of protein structures

The 3D structures of AKT1 (ID: 4EKL) [28, 29] and MAPK1 (ID: 6OPH) [30-32] were retrieved from the Protein Data Bank RCSB (<u>https://www.rcsb.org/</u>). We removed water molecules and co-crystal ligands from the protein molecule by using Discovery Studio software. Next, we used MGL Autodock Tools 1.5.6 software to add hydrogen atoms, optimize polar hydrogens, and Kollman charges. The active region of AKT1 and MAPK1 were located in grid boxes of corresponding sizes ($40A \times 40A \times 40A$) and ($60A \times 60A \times 60A$), the grid center (x,y,z) corresponding (x = 20.918; y = 2.534; z = 16.36) and (x = 5.953, y = 18.56, z = 23.812). This protein

was saved in pdbqt format to prepare for the docking program. The selection of grid box indexes is based on previous publications [32-34].

5.4.2. Preparation of ligands

Based on previous publications, we collected 120 natural compounds in the plant *Zanthoxylum simulans* [11, 18, 21, 25, 36-40]. The ligand structures of these molecules were downloaded from the PubChem database (<u>https://pubchem.ncbi.nlm.nih.gov/</u>) in sdf format. Then we converted them to pdb format using Discovery Studio Visualizer 2021 software. Finally, the ligands were optimized by Avogadro software using Conjugate Gradients method and converted to the pdbqt format using Autodock Tools 1.5.6.

5.4.3. Evaluation of docking results

To evaluate the docking results, the co-crystal ligand after being separated from the protein was redocked to the active site of the target. The process was performed successfully if the root-mean-square deviation (RMSD) value was less than or equal to 1.5 Å. For substances that need docking, their binding ability is assessed through interaction with amino acids, and the interaction energy is calculated by the scoring function of Autodock vina.

5.4.4. Evaluation of Lipinski's rule of five

Lipinski's rule of five was used to study the drug-like and non-drug-like molecules which help to evaluate the potential molecular to become a therapeutic drug. We used the online tool (<u>http://www.scfbio-iitd.res.in/software/drugdesign/lipinski.jsp</u>) and through criteria: molecular mass (MW), high lipophilicity (LogP), hydrogen bond donors (HBD), hydrogen bond acceptors (HBA1); molar refractivity (MR). After selecting the drug-like compounds, we continued to analyze the pharmacokinetic and toxicological parameters to give the final results.

5.4.5. Prediction of ADMET by computational analysis

We used the online tool pkCSM (<u>http://biosig.unimelb.edu.au/pkcsm/prediction</u>) to predict the pharmacokinetic and toxicological properties of the compounds as input data SMILES formulas. Predictive results of pharmacokinetic-toxicological (ADMET) parameters including absorption, distribution, metabolism, elimination, and toxicity of potential compounds were analyzed.

Acknowledgements: This study was funded by University of Medicine and Pharmacy, Vietnam National University Hanoi, with grant number CS.22.02.

Author contributions: Concept – B.T.T.; Design – B.T.T., N.T.T; Supervision – B.T.T.; Resources – L.T.H., T.H.M., V.M.H., D.T.H.K.; Literature Search – L.T.H., T.H.M., V.M.H., D.T.H.K.; Writing – L.T.H., T.H.M., V.M.H., D.T.H.K.; Critical Reviews – B.T.T., N.T.T;

Conflict of interest statement: The authors declared no conflict of interest.

REFERENCES

- [1] Meng X, Zhong J, Liu S, Murray M, Gonzalez-Angulo AM. A new hypothesis for the cancer mechanism. Cancer Metastasis Rev. 2012;31:247-268. <u>https://doi.org/10.1007/s10555-011-9342-8</u>
- [2] Husain S, Szabo I, Pai R, Soreghan B, Jones M, Tarnawski A. MAPK (ERK2) kinase a key target for NSAIDs-induced inhibition of gastric cancer cell proliferation and growth. Life Sci. 2001;69(25-26):3045-3054. https://doi.org/10.1016/S0016-5085%2808%2983299-4
- [3] Qu JL, Qu XJ, Zhao MF, Teng YE, Zhang Y, Hou KZ, Jiang Y-H, Yang X-H, Liu Y-P. Gastric cancer exosomes promote tumour cell proliferation through PI3K/Akt and MAPK/ERK activation. Dig Liver Dis. 2009;41(12):875-880. https://doi.org/10.1016/j.dld.2009.04.006
- [4] Zhou W, Wu J, Zhu Y, Meng Z, Liu X, Liu S, Ni M, Jia S, Zhang J, Guo S. Study on the mechanisms of compound Kushen injection for the treatment of gastric cancer based on network pharmacology. BMC Complement Med Ther. 2020;20:6. <u>https://doi.org/10.1186/s12906-019-2787-y</u>
- [5] Han Z, Wu K, Shen H, Li C, Han S, Hong L, Shi Y, Liu N, Guo C, Xue Y, Qiao T. Akt1/protein kinase Bα is involved in gastric cancer progression and cell proliferation. Dig Dis Sci. 2008;53:1801-1810. <u>https://doi.org/10.1007/s10620-007-9824-2</u>
- [6] Zhou W, Fu X, Zhang L, Zhang J, Huang X, Lu X, Shen L, Liu B, Liu J, Luo H. The AKT1/NF-kappaB/Notch1/PTEN axis has an important role in chemoresistance of gastric cancer cells. Cell Death Dis. 2013;4(10):1-11. https://doi.org/10.1038/cddis.2013.375
- [7] Almhanna K, Strosberg J, Malafa M. Targeting AKT protein kinase in gastric cancer. Anticancer Res. 2011; 31(12): 4387-4392.

- [8] Nikolaou M, Pavlopoulou A, Georgakilas AG, Kyrodimos E. The challenge of drug resistance in cancer treatment: a current overview. Clin Exp Metastasis. 2018;35:309-318. <u>https://doi.org/10.1007/s10585-018-9903-0</u>
- [9] Jain S, Dwivedi J, Jain PK, Satpathy S, Patra A. Medicinal plants for treatment of cancer: A brief review. Pharmacogn J. 2016;8(2): 87-102. <u>https://doi.org/10.5530/pj.2016.2.1</u>
- [10] Ombito JO. Phytochemistry and pharmacology of the genus Zanthoxylum (Rutaceae): A Review. Nat Prod J. 2021;11(1):21-43. <u>https://doi.org/10.2174/2210315509666191202095924</u>
- [11] Chen I-S, Wu S-J, Leu Y-L, Tsai I-W, Wu T-S. Alkaloids from root bark of Zanthoxylum simulans. Phytochemistry. 1996;42(1):217-219. <u>https://doi.org/10.1016/0031-9422(95)00856-X</u>
- [12] Lim T. Zanthoxylum simulans. In: Edible Medicinal And Non-Medicinal Plants, 2012, pp.904-911. http://dx.doi.org/10.1007/978-94-007-4053-2_105
- [13] Larregieu CA, Benet LZ. Drug discovery and regulatory considerations for improving in silico and in vitro predictions that use Caco-2 as a surrogate for human intestinal permeability measurements. AAPS J. 2013;15(2):483-497. <u>https://doi.org/10.1208/s12248-013-9456-8</u>
- [14] Pham-The H, Cabrera-Pérez MÁ, Nam NH, Castillo-Garit JA, Rasulev B, Le-Thi-Thu H, Casañola-Martin GM. In Silico Assessment of ADME Properties: Advances in Caco-2 Cell Monolayer Permeability Modeling. Curr Top Med Chem. 2018;18(26):2209-2229. <u>https://doi.org/10.2174/1568026619666181130140350</u>
- [15] Wang J-F, Deng Y-H, Yang S-H, Liu Y-Q, Wang Y-H, Pan W-W, Zhou X-J. Characterization and biological evaluation of six new dimeric lignans with an unusual α, β-unsaturated ketone motif from *Zanthoxylum simulans*. Bioorg Med Chem Lett. 2014;24(19):4667-4471. https://doi.org/10.1016/j.bmcl.2014.08.042
- [16] Thu NB, Trung TN, Ha DT, Khoi NM, Hung TV, Hien TT, Yim N-H, Bae K-H. Screening of Vietnamese medicinal plants for cytotoxic activity. Nat Prod Sci. 2010;16(1):43-49.
- [17] Tian Y-Q, Hu D, Zhang Y-L, Zou J, Chen G-L, Guo M-Q. Inhibitors targeting multiple Janus Kinases from *Zanthoxylum simulans* mediate inhibition and apoptosis against gastric cancer cells via the estrogen pathway. Front. Chem. 2022;10: 922110. <u>https://doi.org/10.3389/fchem.2022.922110</u>
- [18] Shwu-Jen W, Ih-Sheng CJP. Alkaloids from *Zanthoxylum simulans*. Phytochemistry. 1993;34(6):1659-1961. https://doi.org/10.1016/S0031-9422(00)90870-7
- [19] Maduka TO, Ikpa CB. Zanthoxylum rhetsa (Roxb.) DC.: A systemic review of its ethnomedicinal properties, phytochemistry and pharmacology. World News Nat. Sci. 2021;37:41-57.
- [20] Alam F, Najum us Saqib Q, Waheed A. Cytotoxic activity of extracts and crude saponins from Zanthoxylum armatum DC. against human breast (MCF-7, MDA-MB-468) and colorectal (Caco-2) cancer cell lines. BMC Complement Altern Med. 2017;17(1):368. <u>https://doi.org/10.1186/s12906-017-1882-1</u>
- [21] Chen IS, Wu SJ, Tsai IL. Chemical and bioactive constituents from Zanthoxylum simulans. J Nat Prod. 1994;57(9):1206-1211. <u>https://doi.org/10.1021/np50111a003</u>
- [22] Chang HS, Lee SJ, Yang CW, Chen IS. Cytotoxic sesquiterpenes from *Magnolia kachirachirai*. Chem Biodivers. 2010;7(11):2737-2747. https://doi.org/10.1002/cbdv.200900418
- [23] Ih-Sheng C, Shwu-Jen W, Yuh-Chwen L, Ian-Lih T, Seki H, Feng-Nien K, Che-Ming T. Dimeric 2-quinolone alkaloid and antiplatelet aggregation constituents of *Zanthoxylum simulans*. Phytochemistry. 1994;36(1):237-239. https://doi.org/10.1016/S0031-9422(00)97045-6
- [24] Sumary DP, Mgina CA, Joseph CC. Isolation and antimicrobial activities of a novel discolornolide and other compounds from *Monanthotaxis discolor*. Nat Prod Res. 2020;34(22):3163-3168. <u>https://doi.org/10.1080/14786419.2018.1553168</u>
- [25] Chen I-S, Tsai I-W, Teng C-M, Chen J-J, Chang Y-L, Ko F-N, Lu MC, Pezzuto JM. Pyranoquinoline alkaloids from *Zanthoxylum simulans*. Phytochemistry. 1997;46(3):525-529. <u>https://doi.org/10.1016/S0031-9422(97)00280-X</u>
- [26] Hu J, Zhang WD, Liu RH, Zhang C, Shen YH, Li HL, Liang MJ, Xu XK. Benzophenanthridine alkaloids from *Zanthoxylum nitidum* (Roxb.) DC, and their analgesic and anti-inflammatory activities. Chem Biodivers. 2006;3(9):990-995. <u>https://doi.org/10.1002/cbdv.200690108</u>
- [27] Orellana EA, Kasinski AL. Sulforhodamine B (SRB) assay in cell culture to investigate cell proliferation. Bio-protocol. 2016;6(21):1-7. <u>https://doi.org/10.21769/BioProtoc.1984</u>
- [28] Qu Y, Yang X, Li J, Zhang S, Li S, Wang M, Zhou L, Wang Z, Lin Z, Yin Y, Liu J, Wang N, Yang Y. Network pharmacology and molecular docking study of Zhishi-Baizhu herb pair in the treatment of gastric cancer. Evid Based Complement Alternat Med. 2021; 2021:2311486. <u>https://doi.org/10.1155/2021/2311486</u>
- [29] Du K, Ma W, Yang C, Zhou Z, Hu S, Tian Y, Zhang H, Ma Y, Jiang X, Zhu H, Liu H, Chen P, Liu Y. Design, synthesis, and cytotoxic activities of isaindigotone derivatives as potential anti-gastric cancer agents. J Enzyme Inhib Med Chem. 2022;37(1):1212-1226. <u>https://doi.org/10.1080/14756366.2022.2065672</u>
- [30] Singh VJ, Sharma B, Chawla PA. Recent developments in mitogen activated protein kinase inhibitors as potential anticancer agents. Bioorg Chem. 2021; 114:105161. <u>https://doi.org/10.1016/j.bioorg.2021.105161</u>
- [31] Lebedev TD, Khabusheva ER, Mareeva SR, Ivanenko KA, Morozov AV, Spirin PV, Rubtsov PM, Snezhkina AV, Kudryavtseva AV, Sorokin MI, Buzdin AA, Prassolov VS. Identification of cell type-specific correlations between ERK activity and cell viability upon treatment with ERK1/2 inhibitors. J Biol Chem. 2022;298(8): :102226. https://doi.org/10.1016/j.jbc.2022.102226
- [32] Song S, Zhou J, Li Y, Liu J, Li J, Shu P. Network pharmacology and experimental verification based research into the effect and mechanism of Aucklandiae Radix-Amomi Fructus against gastric cancer. Sci Rep. 2022;12(1):9401. https://doi.org/10.1038/s41598-022-13223-z

- [33] Gu S, Xue Y, Gao Y, Shen S, Zhang Y, Chen K, Xue S, Pan J, Tang Y, Zhu H. Mechanisms of Indigo naturalis on treating ulcerative colitis explored by GEO gene chips combined with network pharmacology and molecular docking. Sci Rep. 2020;10(1):15204. <u>https://doi.org/10.1038/s41598-020-71030-w</u>
- [34] Tang L, Zhang S, Ji JC, Wang PJ, Zhang M, Feng PM, Gao XL. Identifying the mechanisms of Rosa Roxburghii Tratt on treating gastric cancer: Combining the targetable screening from the cancer genome atlas with network pharmacology. Nat Prod Commun. 2021;16(11). <u>https://doi.org/10.1177/1934578X211059646</u>
- [35] Deyá AD. Anti-cancer properties of benzophenanthridine alkaloids from Zanthoxylum genus-in silico evidence. Rev Latinoam de Quimica. 2022 ; 26-44.
- [36] Ko FN, Chen IS, Wu SJ, Lee LG, Haung TF, Teng CM. Antiplatelet effects of chelerythrine chloride isolated from *Zanthoxylum simulans*. Biochim Biophys Acta. 1990;1052(3):360-365. <u>https://doi.org/10.1016/0167-4889(90)90144-3</u>
- [37] Yang YP, Cheng MJ, Teng CM, Chang YL, Tsai IL, Chen IS. Chemical and anti-platelet constituents from Formosan *Zanthoxylum simulans*. Phytochemistry. 2002;61(5):567-572. <u>https://doi.org/10.1016/S0031-9422(02)00268-6</u>
- [38] Li DX, Liu M, Zhou XJ. A new dimeric lignan from *Zanthoxylum simulans*. Zhongguo Zhong Yao Za Zhi. 2015;40(14):2843-2848.
- [39] Chyau CC, Mau JL, Wu CM. Characteristics of the steam-distilled oil and carbon dioxide extract of *Zanthoxylum simulans* fruits. J Agric Food Chem. 1996;44(4):1096-1099. <u>https://doi.org/10.1021/jf950577d</u>
- [40] Qi H, Wang WX, Dai JL, Zhu L. In vitro anthelmintic activity of *Zanthoxylum simulans* essential oil against *Haemonchus contortus*. Vet Parasitol. 2015;211(3-4):223-227. <u>https://doi.org/10.1016/j.vetpar.2015.05.029</u>